

This item was submitted to [Loughborough's Research Repository](#) by the author.
Items in Figshare are protected by copyright, with all rights reserved, unless otherwise indicated.

Improving the assessment of polluted sites using an integrated bio-physico-chemical monitoring framework

PLEASE CITE THE PUBLISHED VERSION

<https://doi.org/10.1016/j.chemosphere.2021.133344>

PUBLISHER

Elsevier

VERSION

AM (Accepted Manuscript)

PUBLISHER STATEMENT

This paper was accepted for publication in the journal Chemosphere and the definitive published version is available at <https://doi.org/10.1016/j.chemosphere.2021.133344>.

LICENCE

CC BY-NC-ND 4.0

REPOSITORY RECORD

Garg, Anuradha, Brijesh K Yadav, Diganta Das, and Paul Wood. 2021. "Improving the Assessment of Polluted Sites Using an Integrated Bio-physico-chemical Monitoring Framework". Loughborough University. <https://hdl.handle.net/2134/19292120.v1>.

Improving the assessment of polluted sites using an integrated bio-physico-chemical monitoring framework

Anuradha Garg^a, Brijesh K Yadav^{a,*}, Diganta B Das^{b,*}, Paul J Wood^c

^a *Department of Hydrology, Indian Institute of Technology Roorkee, Uttarakhand, India*

^b *Department of Chemical Engineering, Loughborough University, Loughborough, Leicestershire,
LE11 3TU, UK*

^c *Geography and Environment, Loughborough University, Loughborough, Leicestershire, LE11 3TU,
UK*

* Corresponding authors (Emails: brijesh.yadav@hy.iitr.ac.in; d.b.das@lboro.ac.uk)

Abstract

Soil – water pollution resulting from anthropogenic activities is a growing concern internationally. Effective monitoring techniques play a crucial role in the detection, prevention, and remediation of polluted sites. Current pollution monitoring practices in many geographical locations are primarily based on physico–chemical assessments which do not always reflect the potential toxicity of contaminant ‘cocktails’ and harmful chemicals not screened for routinely. Biomonitoring provides a range of sensitive techniques to characterise the eco–toxicological effects of chemical contamination. The bioavailability of contaminants, in addition to their effects on organisms at the molecular, cellular, individual, and community level allows the characterisation of the overall health status of polluted sites and ecosystems. Quantifying bioaccumulation, changes to community structure, faunal morphology, behavioural, and biochemical responses are standard procedures employed in biomonitoring studies in many High-Income Countries (HICs). This review highlights the need to integrate biomonitoring tools alongside physico–chemical monitoring techniques by using ‘effect–based’ tools to provide more holistic information on the ecological impairment of soil–water systems. This paper considers the wider implementation of biomonitoring methods in Low to Middle Income Countries (LMICs) and their significance in pollution investigations and proposes an integrated monitoring framework that can identify toxicity drivers by utilising ‘effect–based’ and ‘risk–based’ monitoring approaches.

Keywords: Biomonitoring, organisms, contaminant, polluted sites, integrated monitoring

1. Introduction

The quality of soil and water resources has declined significantly during the post-industrial period due to pollution from agricultural, urban, and industrial sources (Delmail, 2014; Chandrappa and Das, 2021). The soil and water resources of polluted sites may contain highly elevated concentrations of multiple contaminants (e.g., pesticides, petroleum based compounds, dyes, toxic organics, suspended solids and heavy metals) compared to the natural surrounding environment (Mohammadzadeh Pakdel and Peighambardoust, 2018; Ali *et al.*, 2020). The unregulated and uncontrolled discharge of waste into the environment in many regions of the world is largely responsible for the high levels of freshwater and soil–water contamination (Debén *et al.* 2017; García-Seoane *et al.* 2018; Barnett-Itzhaki *et al.* 2018; Mangadze, Dalu, and William Froneman 2019; Ighalo and Adeniyi 2020). Increasing contamination levels disturb and disrupt the ecological functioning of polluted ecosystems. For example, elevated nutrient availability and production in the soil/water environment with resulting effects on inter- and intra-specific interactions among soil microbial communities and freshwater organisms (Beniah Obinna and Ebere, 2019). Depending on the concentration and bioavailability of pollutants, this may result in adverse effects on human health observed via irritation and allergic reactions upon contact, through to chronic diseases or organ failure as a result of long term exposure and, in extreme instances death (Martin and Griswold, 2009; Beniah Obinna and Ebere, 2019). These human health outcomes may occur more frequently and present significant greater challenges within low-income countries due to their limited ability to reduce pollution exposure and limited access to public health facilities for the majority of the population (Brainerd and Menon, 2015; Lavaine, 2015; Wang and Yang, 2016a). In addition, the contamination diminishes the aesthetic quality of the environment through its impact on the odour, colour and, transparency of water as a result of its contact, transport, and deposition of debris, tar, plastic, and other waste (WHO, 2003; T. Zhang *et al.*, 2020). Consequently, effective monitoring represents significant challenges to the management of polluted sites (Behmel *et al.*, 2016; Ali *et al.*, 2020). Effective monitoring requires resource managers to characterise the environmental status and baseline conditions for determining future management, remediation, and restoration activities. Soil–water quality monitoring is undertaken drawing on both long-term records (where available) and the application of standardized measurements to define the quality / health status and temporal dynamics of the site (Bartram and Ballance 1996; Behmel *et al.*, 2016; Bo *et al.* 2017; Chandrappa and Das, 2021). The aim of these activities is to develop a standardized long-term monitoring strategy that is spatially

distributed and that is able to assess the effects of ongoing / current activities that influence the current soil-water quality status (Bartram and Ballance 1996; Bo et al. 2017).

The assessment of polluted sites includes analysing the quality status of land (including soil) and water resources. Water quality testing is the most widely undertaken approach, although soil monitoring is rarely undertaken in association with this due to financial constraints (especially in low-to-middle income countries), and to avoid potential duplication of effort (Duarte *et al.*, 2018; Huang *et al.*, 2019). Water quality monitoring (WQM) programmes are typically designed to provide site specific, relevant, precise, and reliable information regarding the status of a site over space and time. A common challenge in soil and water monitoring programmes is the poor spatial coverage which frequently results in the extrapolation of results (Bartram and Ballance 1996; Harmancioglu et al. 1999; Behmel *et al.*, 2016). Other challenges include the inappropriate selection of monitoring sites, inadequate sampling frequency and the limited number of parameters considered. In order to address these challenges, alongside ongoing scientific advances, there is a need to consider new monitoring approaches, technologies, and sensors (Winkler et al. 2008; Winkelbauer et al. 2014; Altenburger *et al.*, 2019) to accurately characterise the overall health status of a site. The current monitoring ‘toolbox’ available to scientists and regulators needs to be updated to minimize the inaccuracy associated with the pressures and effects of soil-water pollution of individual and networks of sites at the river basin scale (Carvalho *et al.*, 2019).

Physico-chemical analysis represents the foundation of historic soil and water quality monitoring. The majority of low-to-middle income countries (LMIC) in Asia, Africa, Latin America and Middle-East still follow this conventional practise (Mangadze *et al.*, 2019; El Sayed et al. 2020). Although chemical assessments are essential, they are limited by not considering the effects of 1) emerging contaminants 2) ‘contaminant cocktails’ or pollution mixtures 3) different bioavailability or concentrations of contaminants in soil, water, sediments or biota, and 4) eco-toxicity of chemical substances not routinely screened (Villares et al. 2001; Gosavi *et al.*, 2004; Amiard-Triquet et al. 2015; Schöne & Krause, 2016; Prabhakaran *et al.*, 2017). It also does not take in account the impact of chemical pollution on the functioning and survival of biological communities, which are potentially important as early warning indicators for human health risks (Gosavi *et al.*, 2004; Milinkovitch *et al.*, 2019). Chemical analysis alone, cannot characterise the ecological health of a system at different spatial and temporal resolutions, and may lead to inadequate screening of polluted sites (USEPA 2005; Zhou et al. 2008; Schöne & Krause, 2016).

In view of these challenges, this review examines the opportunities for improving current practices within LMICs through incorporation of biomonitoring into conventional environmental monitoring approaches (Delmail, 2014; Altenburger *et al.*, 2019). This paper proposes utilizing available state of the art monitoring technologies but also emphasises the importance of fully integrated monitoring frameworks for thorough assessment of polluted sites (specifically integrating biological, chemical, and physical approached). An integrated approach to characterise the physical status, chemical concentrations, and biological effects of pollution will maximise benefits, reduce the risks to human health and ultimately make most effective use of the resources available. The authors propose that such an integrated programme should be employed globally, with appropriate adjustments based on the local geographical conditions and constraints. The highlighted biomonitoring techniques are not proposed as a substitute for physico-chemical monitoring, but to complement the existing tools for identification and confirmation of the contaminants of interest. Scientists have emphasized the need for integrated monitoring tools in order to establish cause-effect relationships over many years (e.g., Reineke *et al.*, 2002; Delmail, 2014; Altenburger *et al.*, 2019; Brack *et al.*, 2017; Milinkovitch *et al.*, 2019), however, the lack of standardised frameworks and clearly-defined methodologies has impeded their wider application in the field.

Specific contaminants, like hydrophobic substances, are typically persistent, bioaccumulative and toxic in water and are poorly monitored in most localities due to their high spatial variability in water

bodies (Brack *et al.*, 2017). In such cases, the toxicity to organisms associated with the chemical exposure provides the best assessment approach rather than a complete physical and chemical analysis of the ‘whole waterbody’. Similarly, polar molecular substances experience high temporal variations in concentrations in water, requiring frequent sampling to provide clear information on the contamination level (Brack *et al.*, 2017). Bioaccumulation and toxicity monitoring facilitates the detection and quantification of such chemicals (Booij *et al.*, 2016; Brack *et al.*, 2017). An integrated approach may therefore, potentially reduce the overall sampling frequency by focussing on organisms via passive sampling and toxicity profiling by prioritizing specific sampling locations, identifying hot spots and establishing ‘cause and effect’ relationships (Brack *et al.*, 2017). Thus, biomonitoring potentially provides cost-effective solutions, which may be especially beneficial within low-to-middle income countries (LMIC) where frequent sampling alongside highly sensitive (and economically costly) chemical analysis is challenging (Japitana *et al.*, 2018). The availability of sophisticated analytical instruments for a whole range of emerging compounds (e.g., surfactants, pesticides and anti-inflammatory drugs) and their active metabolites, is more challenging for LMICs (Schöne and Krause, 2016; Prabhakaran *et al.*, 2017; Calvo-Flores *et al.* 2018; Gogoi *et al.* 2018; Hybská *et al.* 2020). Analysing trace, yet toxic, concentrations of substances is a resource intensive and expensive procedure. The world is in indispensable need of monitoring approaches that are both scientifically sound and cost-effective in identifying and predicting the potential consequences to the ecosystem and human health (Zalewski, 2015; Brack *et al.*, 2017; Prabhakaran *et al.*, 2017).

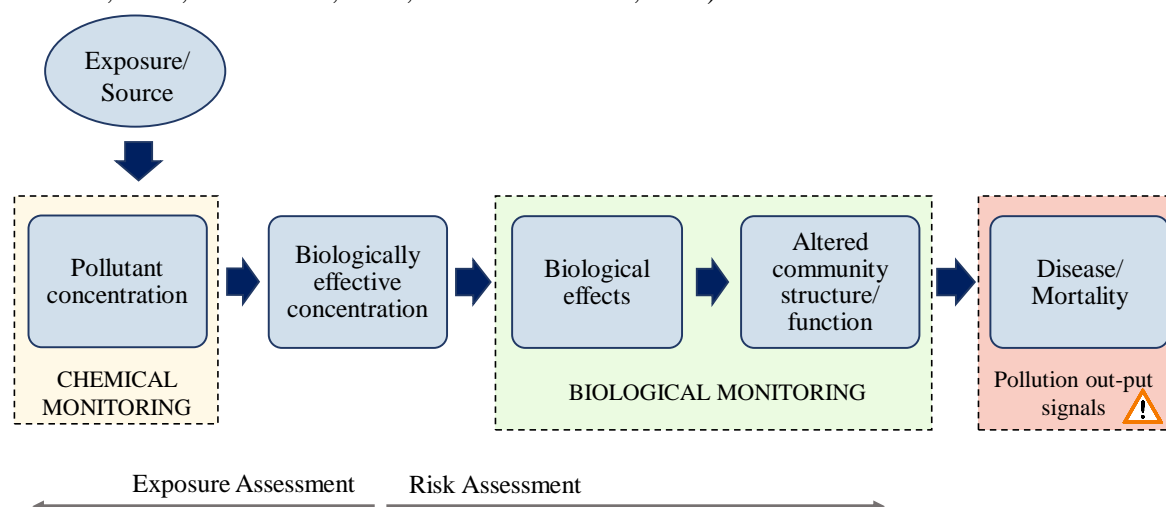


Figure 1: Pollutant pathways from physical exposure, traditional chemical monitoring and biological monitoring of pollutants to identify potential biological effect and risks to human health via disease and mortality.

The passage of pollutants through different components of the monitoring pathway after being released from their source are illustrated in Figure 1. Monitoring systems globally follow a receptor-oriented approach where pollution episodes may be directly related to environmental, ecological, or human health effects. Negative human and ecosystem responses to poor physico-chemical quality requires remediation actions within the affected area(s). Changes in the species composition of a biological community, diversity and mortality represent ‘late indicators’ that should be identified and addressed earlier wherever possible. The scientific community are consistently developing and advancing monitoring tools that can facilitate timely ‘prevention’ rather than ‘remediation’ plans (Milinkovitch *et al.*, 2019). Incorporating ‘early-stage’ biomonitoring (effects on species and community structure and function) within monitoring plans facilitates early intervention that will avoid future stress on the ecosystem and reduce risk to human health (Bolognesi 2003; Barnett-Itzhaki *et al.* 2018; Mangadze *et al.*, 2019). This will lead to the establishment of cost-effective ‘prevention’ plans rather than expensive ‘remediation’ measures.

Accumulation of pollutants into biomass via environmental pathways or the food chain, is critical for the estimation of total pollution loading and lethal/sub-lethal effects (Lovett Doust *et al.* 1994; Adu-manu *et al.*, 2017). Until now, biomonitoring experiments have primarily been conducted as part of independent scientific research by biologists, ecologists, and/or environmental scientists and have not been widely used in field studies within LMICs due to lack of standardised methods and clearly defined protocols (Debén *et al.*, 2017; Altenburger *et al.*, 2019). European countries (e.g., Belgium, France, Italy, UK, Germany and Portugal) along with the USA, Australia, New Zealand, and Canada regularly undertake biomonitoring to assess and monitor the ecological status of surface water bodies (Vanderpoorten 1999; Vincent, Lawlor, and Tipping 2001; Kapfer *et al.* 2012; Gecheva *et al.* 2015; Guareschi, Laini, and Sánchez-Montoya 2017; Pratas *et al.* 2017; Favas *et al.* 2018). This approach has been demonstrated to be especially effective if the wider ecosystem is also regularly monitored along with regular physico-chemical analysis. Milinkovitch *et al.*, (2019) highlighted that the alterations in ecological parameters (e.g., species abundance and diversity) may occur due to chemical stressors and/or other factors including temperature, resource availability and salinity. Ecological evaluation alone cannot identify the primary factors generating ecosystem level impacts (Schiedek *et al.*, 2007; Thrush *et al.*, 2008; Moe *et al.*, 2013). As a result, an ‘effects’ based approach is necessary to address pollution (Brack *et al.*, 2017; Vethaak *et al.*, 2017; Altenburger *et al.*, 2019; Milinkovitch *et al.*, 2019). The EU Water Framework Directive (WFD) requires biomonitoring and physio-chemical monitoring techniques to be applied in all surface water quality monitoring programmes focussed at the ecosystem level. This paper advocates the need to constantly reconsider and update existing programmes, including those utilized as part of the WFD, and especially the establishment of new integrated programmes for LMICs. The areas where the proposed framework outlined and advocated in this paper may contribute in comparison to the existing approaches utilised within the EU WFD are outline in Table 1. The approach will provide benefits to both more high income nations (in improving and reviewing current strategies) and LMIC economies (in adopting new and cost-effective monitoring tools).

Table 1: The proposed framework for pollution monitoring of soil-water resources compared to the European Union Water Framework Directive (EUWFD)

	Proposed Framework	EU WFD
Biomonitoring tool	Ecosystem Bioaccumulation Toxicity	Ecosystem
Target sites	Water resources of polluted sites	All surface waters
Application	Internationally (especially LMICs)	European countries

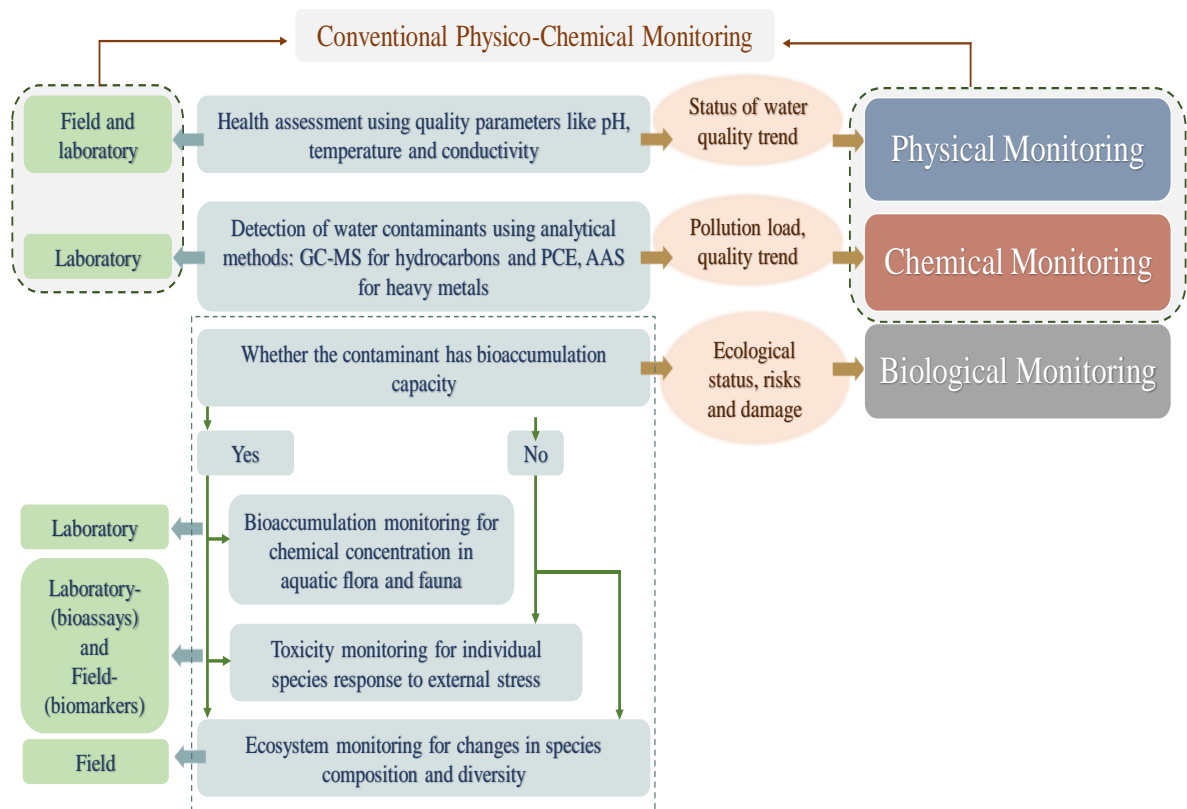


Figure 2: Physical, chemical, and biological parameters and monitoring techniques used in characterizing polluted sites along with their target objectives

A representation of the physical, chemical and biological monitoring objectives and measurements procedures advocated are outline in Figure 2. While physico-chemical monitoring combines approaches, biological parameters rarely form the part of the assessment methodology in most LMICs (Debén *et al.*, 2017). Practically, a single methodological approach cannot fulfil all monitoring requirements of impaired sites with a history of multiple pollution episodes (Altenburger *et al.*, 2019). Combining approaches and pathways will provide a mechanism for improving the overall quality and coverage of environmental monitoring programmes. Within LMICs, comprehensive soil and water quality monitoring is still at a relatively nascent stage of development (McBratney, Field and Koch, 2014; Bünemann *et al.*, 2018; Odountan *et al.*, 2019), and needs to be more cost-effective for its wider implementation. Generating reliable information within the confines of an economically feasible monitoring process is crucial for regions where financial constraints may limit the uptake of technological developments. Attempting to transfer monitoring frameworks from more economically developed regions (e.g. like those within the European Water Framework Directive), where environmental, social and economic conditions are different, may result in inadequately characterising data of local water quality issues (Behmel *et al.*, 2016). The process of data collection, analysis, and management of water quality data demands significant financial resources, professional expertise, equipment, and laboratories. These resources are limited or not widely available in LMICs and requires a greater focus on formulating recommendations for WQM pathways within the constraints of limited economic resources. Although biomonitoring is well established in some HICs, there is only an emerging body of research in LMICs, it has not been theoretically and practically adapted to addressing these limitations. This deficiency in literature from LMICs justifies the need to shift the focus towards a novel bio-integrated approach as proposed herein. This paper highlights the potential value and

significance of biomonitoring in addressing these issues with reference to polluted site subjected, but not limited to heavy metals, organics, hydrocarbon oils, and emerging pollutants. It advocates the use of different organizational levels of responses (e.g., biomolecular, morphological and diversity) of organisms and communities to complement physical and chemical analyses to be better prepared to manage and mitigate environmental perturbations / pollution where it occurs. This paper therefore attempts to improve knowledge on the application of standardized bio-integrated monitoring methodologies that can be applied across diverse geographical regions globally. The paper specifically contributes to the literature by advocating greater uptake of the approach within LMICs, where the advantages of biomonitoring are not currently explored to improve their conventional polluted site monitoring methods. While this paper is mainly focussed on targeting polluted water sites, similar approach can be applied and may be beneficial for polluted soil/land including brownfields where plants and soil microbial communities may act as biomonitors/bioindicators (Saunier *et al.*, 2013; Dadea *et al.*, 2017).

2. Biomonitoring

The term ‘biomonitoring’ can have different meanings depending on the disciplinary background of the individual(s). In the broadest sense, any biological measurement that aims to measure, protect and preserve natural ecosystems can be called biomonitoring (Zwart, 1995). The selection of biological monitoring approaches may be determined by the time, scale, stressor, and sensitivity of the measurement required. It can range from concentration measurement of pollutants within tissues to quantifying their large-scale ecological community impacts. Ecohydrology, which studies the two-way interaction between water and biota, can be fundamentally associated with biomonitoring as it can help in first characterising and subsequently achieving environmentally sustainable quality goals (Prabhakaran *et al.*, 2017). Biological responses depend on the magnitude, frequency and duration of exposure to stressors / contaminants. The reaction to pollutants may occur at three different levels: 1) interaction of stressor with organisms cells, 2) activation of cells responding to the stressors, and 3) adaptive response to maintain functioning (failure to adapt may result in death of the cell / organism) (Piña and Barata, 2011). The contaminant dose is an equally important factor as lower levels may result in adaptation while higher levels may result in acute toxicity and physiological responses by the biota, that may ultimately lead to death (Piña and Barata 2011; Amiard-Triquet *et al.* 2015).

2.1 Types of Biomonitoring

Biomonitoring of effects, and risk assessment in relation to chemical exposure to contaminants, follows a series of distinct methodologies developed over many years (Amiard-Triquet and Rainbow, 2009). The principal types of biomonitoring methods outlined here may be performed for specific purposes where the methods used will reflect the requirements in terms of sensitivity, organisms considered and the need to apply the results to the wider ecosystem. The results may enable environmental managers, regulators (government/private agencies), scientists, and the potential end-users to allocate resources to determine the most effective restoration and remediation strategies (Amiard-Triquet *et al.* 2015). Three types of biomonitoring are outlined below:

2.1.1 Bioaccumulation monitoring: This form of monitoring quantifies the concentration of pollutants measured within an organism, biological material or specific tissues (Zwart 1995; Schilderman *et al.* 1999; Salánki *et al.* 2003). The individual species should be examined for any accumulation of ‘pollutants’ or environmental markers within their tissues, biomolecules or DNA (Melville and Pulkownik, 2007; Baldantoni *et al.*, 2018; Favas *et al.*, 2018). Human biomonitoring (HBM) has also been widely applied for detecting the health effects of environmental pollutants, where chemicals and their metabolites are directly measured in human tissues and/or body fluids in medical research (e.g., Barnett-Itzhaki *et al.*, 2018). The approach is widely applied in public health studies by identifying

specific risk groups associated with particular contaminants (Bolognesi 2003; Barnett-Itzhaki et al. 2018; Vieira *et al.*, 2019; Zhang *et al.*, 2020).

2.1.2 Toxicity monitoring: This approach requires the organisms' response to external stressors to be directly studied and quantified (Zwart, 1995). The measurements of an organisms' physiological, morphological and biomolecular responses, such as lethal concentration, survivorship (Bonnail, Macías and Osta, 2019), biomass, growth (Hybská *et al.*, 2020), damage to DNA and other genetic markers (Vieira *et al.*, 2019), phytotoxicity, cytotoxicity, genotoxicity, mutagenity (Cavusoglu *et al.*, 2010; Bilal *et al.*, 2016; Artico, Migita and Menezes, 2018; Olusola and Solomon, 2018) and locomotion responses (Salánki *et al.*, 2003) have been utilised in previous research. Experiments may be conducted in bioassays / mesocosms designed on simplifications of the natural environment, to control for the complexities of real-world field conditions. Monitoring the modification to the organisms behaviour provides an early warning of potentially significant ecological disturbances that may follow due to increasing contaminant / pollution levels (Cavusoglu *et al.* 2010; Bilal *et al.* 2016; Li *et al.* 2018; Bonnail *et al.* 2019; Hybská *et al.* 2020).

2.1.3 Ecosystem monitoring: Changes to community composition (taxonomic changes or community functioning) due to environmental disturbances can be studied as part of ecosystem monitoring approaches. These studies typically require long-term monitoring of the study area and specific sites. Determining the ecosystem quality and health can be based on comparisons between variables such as population density, species composition, abundance, diversity or may be based on specifically indices / metrics developed to characterise individual stressors / contaminants (Zwart 1995; Clark and Clements 2006; Hering *et al.* 2006; Li *et al.* 2010; Delmail 2014; Niba and Sakwe 2018).

A comparative overview of the three principal forms of biomonitoring that may be performed by regulatory, industrial, or academic organizations is outlined in the Figure 3. Bioaccumulation reflects the interaction between the polluting compound and biota via its incorporation within tissue(s). Its application typically depends on the pollutant's properties, bioaccumulation potential, and the biological factors that determine the fate of pollutants within the food chain / web (Amiard-Triquet and Rainbow, 2009). Within this form of biomonitoring, it is crucial to analyse the differences between a contaminants' concentration within both water and soil compared with the bioavailable concentrations within the organism(s). The fraction of the available pollutant concentration entering the food chain may result in significant changes to biological material (Yadav *et al.* 2011). Low sub-lethal concentrations may cause chronic diseases, while lethal concentrations will kill most biota (Specziár 2002; Salánki *et al.* 2003).

Toxicity monitoring is a growing area of eco-hydrological research, especially where accurate monitoring of polluted waters is a major concern and where understanding temporal variability of effects is required (e.g., under controlled laboratory exposure over set time-periods). Depending on the type of pollution and biological variable(s) under investigation, monitoring can be performed in the laboratory or *in-situ* in the field if appropriate control measure can be put in place. Experiments are typically conducted in bioassays in the laboratory, while the field samples may be collected over space and/or time with respect to clearly identified biomarkers. Laboratory based investigation may employ smaller sample sizes than those collected in the field to observe responses to pollution stress compared to non-polluted (control) bioassays under closely monitored experimental conditions.

Changes in the survivorship or growth of individuals within a population or community in response to the input of pollutants forms the basis for ecosystem monitoring. The endpoints of pollution monitoring may be based on the measures of survivorship, growth and reproduction potential, which ultimately lead to changes in the community composition and population sizes. The biological communities inhabiting polluted sites (compared to unpolluted / unstressed sites) provide evidence of the contamination's effect at a polluted site. The biota act as continuous monitors (over their entire life span) of the conditions they experience and can be directly related to instantaneous chemical analysis

(Zwart 1995; Dalu and Froneman 2016). Considering the wide range of pollutants, this approach enables the study of the ‘cocktail effect’ (additive, reductive or synergistic) of pollution mixtures (Fu *et al.*, 2018) although in many instances the nature of the ‘cocktail’ remains unknown.

While ecosystem monitoring facilitates the assessment of ‘risks’, bioaccumulation and toxicity monitoring allows the quantification of pollution ‘effects’. The latter two methodological approaches provide early indicators that bridge the information obtained by chemical analysis and ecosystem monitoring. The three levels of biomonitoring together deliver maximum benefits when undertaken in an integrated solution-oriented framework of polluted sites.

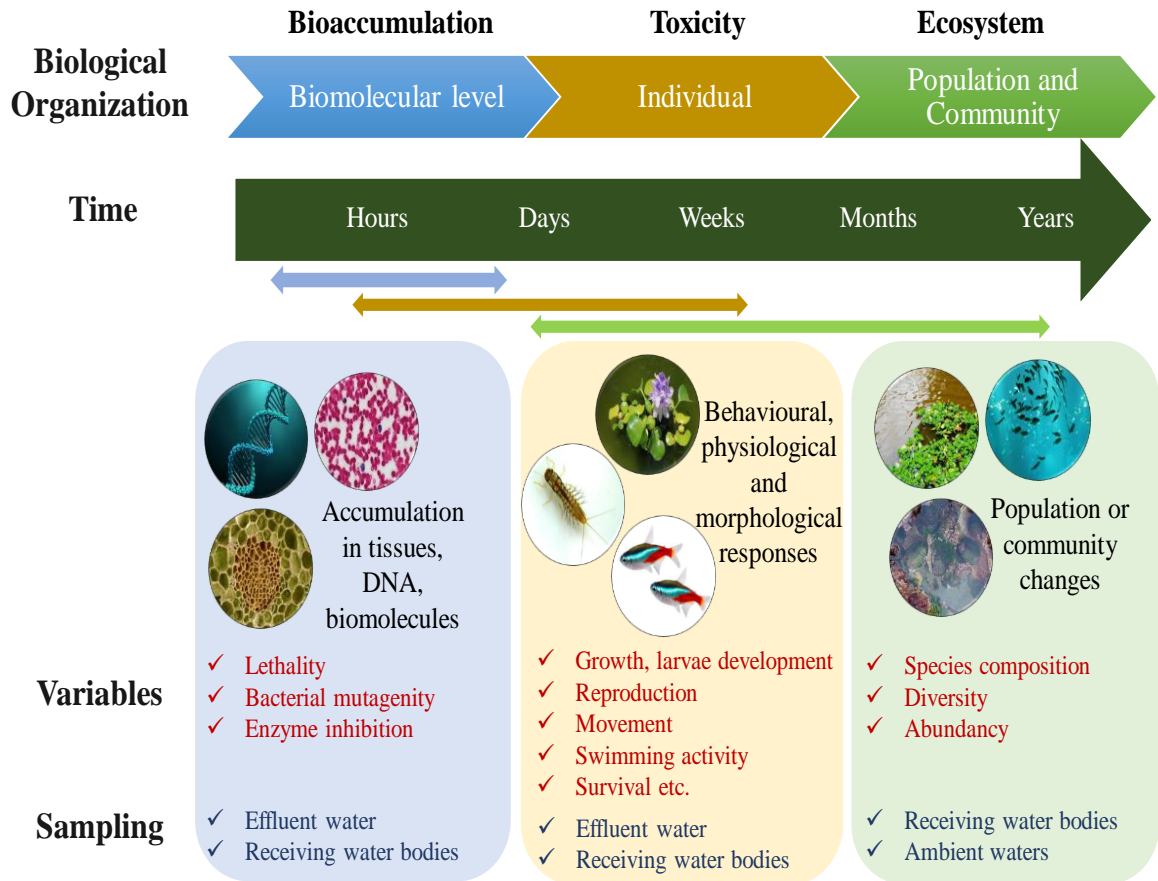


Figure 3: Schematic representation of the three types of biomonitoring techniques widely employed internationally: Bioaccumulation, Toxicity and Ecosystem monitoring. Indicating the level of biological organization, time required for study / analysis, typical types of variables considered, and sampling approach required to implement monitoring/

2.2 Biomonitoring, Biomarkers and Bioindicators

Many scientists describe ‘biomonitors’ and ‘bioindicators’ as the same entity, i.e., a species or group of organisms reflecting the surrounding environment / abiotic conditions, including stressors and pollutants (Rainbow, 1995; Prabhakaran *et al.*, 2017; Samsi *et al.*, 2017). However, the definitions have evolved over time. The biotic species studied can be classified into two types: a) indicator organisms - which display specific tolerances to the environmental/abiotic conditions, and b) bio-accumulators of pollutants. Indicator organisms are typically referred as bioindicators, and accumulators as biomonitors (Phillips and Rainbow 1994; Li *et al.* 2010). Another term used in the literature, ‘biomarkers’, refers to the biological response/characteristics or the presence of (bio)chemical markers whose presence indicates environmental perturbations associated with pollution loading (Celander 2011; Hamza-

Chaffai 2014). The three terms, however, are often used interchangeably despite the differences that are acknowledged by those working directly in the field.

2.3 Selection of species and biomonitoring variables

The success of contaminant biomonitoring depends on the selection of suitable species that form the basis of specific criteria (Hamza-Chaffai 2014; Amiard-Triquet et al. 2015). The choice of variables (both the organisms used and contaminants studied) depends on the context specific circumstances associated with pollutants. The essential features for selection of the most suitable species for biomonitoring have been considered by several authors (Table 1) – Metcalfe-Smith (1994), Zwart (1995), and Amiard-Triquet et al. (2015). The species are primarily selected based on the scientific requirements of the study while the selection of abiotic (including chemicals) variables (parameters) depends on administrative and policy decisions too.

The selection criteria outlined in Table 2 identifies ‘sentinel species’ (indicator taxa / groups of organisms) ideal for the biomonitoring studies. For instance, a species may be selected from a range of ‘sentinel species’ having experienced serious effects (survivorship, growth or population size) due to pollution (Chapman, 2002; Hale and Koprowski, 2018). In aquatic systems, benthic taxa like amphipod Crustacea (e.g., Gammaridae) are important ‘keystone species’. They play a crucial role in detritus processing and comprise an essential part of the aquatic food webs as both shredders of detritus and prey for higher secondary consumers (Amiard-Triquet et al. 2015). These species are sensitive to a wide range of chemical stressors, including heavy metals, organic pollution and hydrocarbons (Von Der Ohe and Liess 2004; Dauvin and Ruellet 2007). Their life cycle and readily recorded responses to abiotic conditions are significant for the scientific understanding of wider community structure (Kunz et al. 2010) and ecosystem functioning. The selection of biomarkers may vary according to the type of contaminant and the environmental setting. Amiard-Triquet *et al.* (2013) utilized several species of micro-algae, nematode worms, bivalves, crustacea, insects, and fishes as biomarkers of tolerance and as early indicators of heavy metal pollution. The list can be extended further by adding phytoplankton, annelid worms, and amphibian species as biomarkers of organic contaminants, including PCBs, PAHs, as well as a range of pharmaceuticals and pesticides (Amiard-Triquet *et al.*, 2013). However, careful selection of biomarkers is required to ensure clear demarcation of the contamination stressor and the influence of confounding factors (non-chemical stressors) (Jemec *et al.* 2010).

Table 2: Criteria for the selection of species and variables for the establishments of biomonitoring programmes. Source: Metcalfe-Smith (1994), Zwart (1995), and Amiard-Triquet et al. (2015)

Criteria for species selection	Criteria for variables selection
<ul style="list-style-type: none"> ▪ Ease of identification ▪ Sedentary nature for representativeness of spatial extent of pollution at the site ▪ Sufficient population size ▪ Ease of sampling ▪ Longevity as compared to other aquatic species for recording temporal changes in the response ▪ Availability for capture throughout the year ▪ Sensitivity to dose-effect and cause-effect relationships ▪ Representativeness towards the environmental conditions 	<ul style="list-style-type: none"> ▪ Ecological and environmental value of the information ▪ Representativeness of response in the studied and other species ▪ Cause (contamination) – effect (variable) relationship specificity ▪ Sensitivity to the pollution stress ▪ Quantifiable response range and rate ▪ Standardization procedure of the measurement method ▪ Applicability to similar sites for comparison ▪ Cost-effectiveness ▪ Retrospection

3. Global history and current research trends in Biomonitoring

Despite the wide application of biomonitoring methods in several HIC regions of the world, it is a relatively novel concept for some low- and middle-income countries (LMICs). European countries including Czech Republic, Italy, the Netherlands, Germany, Sweden, UK, France, Poland, Slovakia, Denmark, Italy Canada, the US, Australia and Asian countries like South Korea, Japan and China have been at the forefront of using biomonitoring techniques and methodologies to define the ecological health status of aquatic waterbodies (Clarke *et al.*, 2006; Hering *et al.*, 2006; Buss *et al.*, 2015). Some regions (e.g., European Union member countries) have developed standardized protocols to monitor and assess the biological status, patterns, and trends in the health / quality of surface waters. These protocols define standard procedures for sampling, collection and identification of biological taxa (Birk *et al.*, 2012a). The regions with a longest history on biomonitoring methods (e.g. Europe and the USA) also have some locally developed small scale biomonitoring programs specific to their geographical location or ecosystem type (Birk *et al.*, 2012b; Buss *et al.*, 2015). Legislation like the EU Water Framework Directive (WFD) in Europe aims to maintain the ecological health of surface water bodies. Similarly, programmes like Australian River Assessment System (AUSRIVAS), Canadian Aquatic Biomonitoring Network (CABIN) and USEPA's National Aquatic Resources Survey (NARS; previously called EMAP) undertake routine biological monitoring to help maintain aquatic ecosystem health. The most commonly and widely used biological indicator group of organisms are macroinvertebrates, followed by fish, bacteria, and algae (Morse *et al.*, 2007; Buss *et al.*, 2015). While the methods have been widely explored and applied in these areas, some global regions such as Latin America, Africa and parts of Asia (encompassing low and middle income countries LMIC) lack a history of biomonitoring of freshwater ecosystems (Buss *et al.*, 2015). In addition, in many tropical nations around the world, a small number of pilot studies have been undertaken, e.g., National River Health Monitoring Program in South Africa like SASS and MIRAI (Dallas, 1997, 2004; Ollis *et al.*, 2006) and thus, it is imperative to develop standardised methods for biomonitoring in these regions. Field applications of biomonitoring focus only on ecosystem monitoring for the assessment of ecological and human health risks and such practices mean that significant knowledge gaps persist (Brack *et al.*, 2017; Milinkovitch *et al.*, 2019).

Generic surface water monitoring (primarily using chemical and physical approaches) may help identify potential pollution sources within the ecosystems and wider catchments. However, this review highlights the need to incorporate biomonitoring of the effects recorded at polluted sites (e.g., industrial area) on soil-water resources. We emphasize the values of integrated approaches, where chemical monitoring may identify specific pollutants, and biomonitoring quantifies the ecological damage and effects caused by pollutants. Starting from the point of effluent discharge and working downstream to receiving water bodies in a systematic manner would potentially save time and reduce costs of monitoring the entire network. The framework in this paper proposes monitoring (bioaccumulation, toxicity, and ecosystem monitoring of specific waters) which tracks the extent and temporal duration of biological degradation (fate and transport). It proposes a focus on measuring the extent of degradation caused from clearly identified pollution sources, rather than plotting deterioration in ecosystem quality. It also emphasizes the importance of the wider field application of bioaccumulation and toxicity monitoring along with ecosystem monitoring. Global regions with a limited history of biomonitoring applications (especially many LMICs) would gain from the establishment of credible integrated monitoring programmes (encompassing chemical, physical and biological approaches). This would be particularly beneficial for areas with finite economic resources, where nation-wide application of one standard method is not feasible

4. Biomonitoring applications

With the increasing occurrence of environmentally persistent toxic compounds, new emerging contaminants, and mixed ‘cocktails’ of pollutants discharging into waterways, the need for robust, readily available and inexpensive pollution assessment methods has grown. Biomonitoring has provided reliable, efficient, accurate and cost effective information over many years of use (Zwart 1995; De Bisthoven et al. 1998; Jemec et al. 2008; Oost et al. 2016; Prabhakaran et al. 2017). The biotic responses of ‘sentinel species’ or ‘taxa’ in the form of pollutant uptake, (Lovett-doust et al. 1994; Meador et al. 1995; Zuykov et al. 2013; Prabhakaran et al. 2017), accumulation (Lee and Wang 2001; Lovett-doust and Lovett-doust 2001; Liu et al. 2016; Nascimento et al. 2018; Vieira et al. 2019), and lethality (Altinok and Capkin, 2007; Paulino *et al.*, 2014; Vieira *et al.*, 2019) have been used to quantify and characterise the wider health of pollutes sites and ecosystems. Examination of the literature indicates that biomonitoring methods are categorized in two types: active and passive. Passive monitoring is performed by collecting ‘resident species’, those organisms naturally inhabiting and growing in the area. In contrast active monitoring, may require the study of organisms transported to the laboratory- although some studies have been undertaken in the field (García-Seoane, Fernández, et al. 2018; Vieira et al. 2019). Passive techniques are more frequently utilized as they provide greater insights to the spatial distribution and extent of bioavailable pollutants under real world conditions (García-Seoane, Aboal, *et al.*, 2018). In active monitoring, a well-studied organism can be exposed to the conditions present or to the specific compound / chemical present at the target (polluted) site for specified periods of time. Comparisons between multiple bio-monitors or bio-indicators can produce statistically powerful results. Comparisons can also be made by using the same species for a range of different pollutants to analyse their bioavailable concentrations or interactions when multiple compounds are present in the environment (Rainbow 1995; Lee and Wang 2001; Nascimento et al. 2018). Examples of previous studies utilising biomonitoring techniques to a range of pollutants present in the natural environment are presented in Table 2.

Table 3: Recent studies on biomonitoring techniques using plants and animal biological indicator species.

Biomonitoring Technique	Bioindicator Species	Pollutant(s)	Biological/Ecological parameter	Reference	Coupled with Physical /chemical monitoring
Bioaccumulation	<i>Konosirus punctatus</i> , <i>Mugil cephalus</i>	Microplastics	Indigestion by fish	Zhang et al. (2020)	Yes
Toxicity	<i>Daphnia magna</i> , <i>Allium cepa</i> , <i>Lemna minor</i>	Waste Water Treatment Plant	Immobilisation in <i>Daphnia</i> , Biomass, growth, and leaf count in <i>Allium cepa</i> and <i>Lemna minor</i>	Hybská et al. (2020)	No
Bioaccumulation and Toxicity	<i>Corbicula fluminea</i>	As, Cd, Cu, Cr, Co, Fe, Mn, Pb, Ni, Sb, Se, Zn	Lethality, Survival, Accumulation in soft tissue	Bonnail et al. (2019)	Yes

Bioaccumulation and Toxicity (Active Monitoring)	<i>Prochilodus lineatus</i>	Organochlorine pesticides (OCPs), Trace metals (Cu, Cr, Cd, Pb, Ni, Zn), Pesticides Macronutrients	Biotransformation and antioxidant enzymes, oxidative damages, DNA damages and liver histopathology	Vieira et al. (2019)	Yes
Bioaccumulation	<i>Apium nodiflorum</i> , <i>Potamogeton pectinatus</i>	micronutrients and toxic elements	Bioconcentration in roots and shoots	Baldantoni et al. (2018)	Yes
Toxicity	<i>Allium cepa</i>	Coal contaminants	Phytotoxicity, cytotoxicity and genotoxicity	Artico et al. (2018)	Yes
Toxicity	<i>Allium cepa</i>	Pharmaceutical effluents	Phytotoxicity, cytotoxicity and genotoxicity	Olusola and Solomon (2018)	Yes
Toxicity	Zebrafish Larvae species	Waste Water Treatment Plant	Change in heartbeat rate, Survival response etc.	Li et al. (2018)	No
Bioaccumulation	<i>Fontinalis squamosa</i> , <i>Brachythecium rivulare</i> , <i>Platyhyphnidium riparioides</i> , <i>Thamnobryum alopecurum</i> , <i>Lemanea fluviatilis</i> (Bryophytes)	46 elements including heavy metals	Phytoaccumulation	Favas et al. (2018)	Yes
Ecosystem	Benthic macroinvertebrates (multiple species)	-	Species abundance, richness, trend	Niba and Sakwe (2018)	Yes
Bioaccumulation	<i>Corbicula fluminea</i>	Lanthanides	Accumulation in soft tissue	Bonnail et al. (2017)	No
Bioaccumulation	<i>Posidonia oceanica</i> , <i>Cymodocea nodosa</i> , <i>Phragmites australis</i> , <i>Arundo donax</i> , <i>Typha domingensis</i> , <i>Apium nodiflorum</i> , and <i>Nasturtium officinale</i>	Heavy metals	Bioconcentration in roots and leaves	Bonanno et al. (2017)	No
Toxicity	<i>Triticum aestivum</i>	Textile effluent	Phytotoxicity and cytotoxicity	Bilal et al. (2016)	No

Bioaccumulation	<i>Fontinalis antipyretica</i> , <i>Sphagnum denticulatum</i>	Heavy metals	Accumulation in moss	Debén et al. (2016)	No
Toxicity	<i>Vicia faba</i>	Petroleum refinery effluent	Cytotoxicity, genotoxicity and mutagenicity	Cavusoglu et al. (2010)	Yes
Bioaccumulation	<i>Caloglossa leprieurii</i> , <i>Catenella nipae</i> , <i>Bostrychia</i> sp.	Heavy metals	Concentration factors in algae	Melville and Pulkownik (2007)	Yes
Bioaccumulation and toxicity	<i>Lymnaea stagnalis</i> L.	Heavy metals	Accumulation in gills, muscles, liver, locomotion pattern, effect on resting behaviour	Salánki et al. (2003)	No
Bioaccumulation	<i>Orconectus limosus</i> (Crayfish)	Heavy metals (Cd, Zn, Cu, Pb) PCBs, PAHs, and Organochlorine pesticides (DDT, DDE)	Accumulation in hepatopancreatic tissue	Schilderman et al. (1999)	Yes

439

440 Early research on bio-accumulation and monitoring largely focussed on quantifying heavy metals
441 in plants and animals (Mathur and Yadav, 2009). Macrophytes are popular taxa for bio-monitoring
442 purposes because they can accumulate significant pollutant concentrations in various body parts (e.g.,
443 roots, shoots and leaves). Baldantoni et al. (2018), for example, observed bio-concentration of micro-
444 nutrients (Ca, K, Mg, P), micronutrients (Cu, Fe, Mn, Na, Ni, Zn), and toxic elements (Cd, Cr, Pb, V)
445 in the roots and shoots of the aquatic macrophytes, *Apium nodiflorum* and *Potamogeton pectinatus*. The
446 variation in concentrations of elements at different sites helped to establish a correlation with potential
447 agricultural, urban, or industrial pollution sources. The concentrations of heavy metals within five
448 different species of bryophytes was also studied by Favas et al. (2018) to highlight species specific
449 bioaccumulation characteristics. Further research by Bonanno et al. (2017), calculated the bio-
450 concentration of heavy metals in the roots and leaves of two seagrasses *Posidonia oceanica* and
451 *Cymodocea nodosa*, and in five wetland macrophytes *Phragmites australis*, *Arundo donax*, *Typha*
452 *domingensis*, *Apium nodiflorum*, and *Nasturtium officinale*. While heavy metal accumulation is mainly
453 studied in macrophytes, oils and organic chemicals bio-accumulation has been studied in faunal species.
454 Contemporary research is using biological indicators to examine microplastics, pesticides and a wide
455 range of heavy metals (e.g., Bonnail et al. 2019; Vieira et al. 2019; Zhang et al. 2020).

456 Toxicity monitoring is widely applied using fish, macroinvertebrates, bryophytes and macrophytes
457 (Li et al. 2018; Bonnail et al. 2019; Hybská et al. 2020; Zhang et al. 2020). Ecosystem monitoring is
458 comparatively less popular in terms of toxicity and sensitivity but has the ability to demonstrate the
459 ecological significance and effects in the natural environment. The technique requires sampling of
460 multiple sites simultaneously (Delmail, 2014) or periodic sampling of the same polluted sites in
461 association with appropriate non-polluted control sites (Niba and Sakwe, 2018) for ongoing tracking of
462 changes in the ecosystem properties (e.g., community structure or biodiversity). Biomonitoring of
463 freshwater aquatic ecosystems widely utilizes benthic macroinvertebrates (Carter et al. 2006; Mathers
464 et al. 2016; Bo et al. 2017). Amphipod crustaceans from the family Gammaridae are among the most
465 widely studied and utilized group of macroinvertebrates used to investigate pollution for more than 90
466 years (e.g., Amiard-Triquet et al. 2015). More than 20 species of *Gammarus* have been utilised in eco-

toxicological studies of aquatic systems. The species are suitable for both laboratory (bioassays) and field toxicity (biomarkers) studies (Kunz et al. 2010); with freshwater species *Gammarus pulex* being the most widely utilised (Kunz et al. 2010; Gerhardt et al. 2012; Besse et al. 2013; Lebrun et al. 2015; Ciliberti et al. 2017; Fu et al. 2018; Shahid et al. 2018; Tatar et al. 2018; Serdar 2019; Serdar et al. 2019; Lebrun and Gismondi 2020). The group of amphipod shrimp (*Gammarus spp.*) are also notable as they have also been used to assess the effects of varying concentrations of chemicals associated with sediments (e.g., Costa et al. 2005; Neuparth et al. 2005; Gaskell et al. 2007). In eco-toxicological studies, population endpoints can be measured in terms of survivorship (mortality), the population structure (age / size classes), density, and interaction with other species (Kunz et al. 2010). Their feeding activities, growth, size, fecundity, locomotion, and survival can all be effectively recorded and used as indicators of chemical / abiotic stressors (Gerhardt 1995; Kunz et al. 2010). Among plant species, macrophytes such as *Eichhornia crassipes* (water hyacinth), bryophytes such as *Brachytheceum rivulare* and *Thamnobryum alopecurum*, and seagrass species including *Posidonia oceanica* and *Cymodocea nodosa* have all been widely used to monitor the accumulation of chemical compounds and their effects on morphology, growth, size, density and physiological functioning (Romero et al. 2006; Bonanno, Borg, and Di Martino 2017; Favas et al. 2018).

5. Example of Biomonitoring Method

In order to demonstrate the principles of biomonitoring, we measured the effects of a species respond to a low-level exposure of a model contaminant. The response of benthic macroinvertebrates *Gammarus pulex* (Crustacea, Amphipoda) under the presence of mild non-aqueous phase liquid (mineral oil) were measured in terms of changes to movement and survivorship. *G. pulex* were collected from Burleigh Brook on the campus of Loughborough University (UK) in a pond net (mesh size = 1mm) using the kick-sampling method. The collected specimens were carefully transferred into containers of stream water and transferred to the laboratory for acclimatization in an aerated mesocosm at 23°C prior to the experiment. This bioassay study used a sandy soil medium with a particle size range of 100 – 250 µm with bulk porosity of 0.25 and hydraulic conductivity of 0.023 cm/s. Mesocosms were set up by taking varying volumetric proportions of soil – water – mineral oil mixture. A bulk density of 1.59 g/ml was maintained initially in all experimental units, which were later supplemented with 150 ml water in order to create the media for the species to be observed within. The treatment mesocosms were spiked with mineral oil to achieve 0 to 25% oil – water mixture. Each mesocosm contained five *G. pulex* and were run simultaneously with four replicates.

Observations of the behaviour and movement of individuals during the experiment indicated that when individuals came into contact with the mineral oil their buoyancy was increased. The results representing the average number of *Gammarus* individuals floating near the water surface was plotted at the end of 24 h study period for each of the treatments (Figure 4). The results and response observed clearly indicated an increasing number of individuals close to the water surface with an increasing ratio of mineral oil – water emulsion (Treatment 1 - 0% mineral oil / control – treatment 7 - 25% mineral oil). The greater number of individuals recorded displaying modified movement due to increased buoyancy were recorded for treatment 6 (20% Mineral Oil: 80% Water). Within Treatment 6, an average of 50% of individuals displayed changes in their movement pattern and were located close to the water surface at end of the experiment. Almost all individuals were alive at the end of the experiment (survivorship was > 90%). These observations illustrate the effects of an oil-based contaminant and change in the movement and behaviour of *G. pulex*. This species typically inhabits river bed sediments (benthic sediments) where it seeks refuge between sediment clasts. The behavioural shift with the species being confined to the surface layers of the mesocosm would make individuals more vulnerable to predation by the higher-level organisms, especially predatory fish.

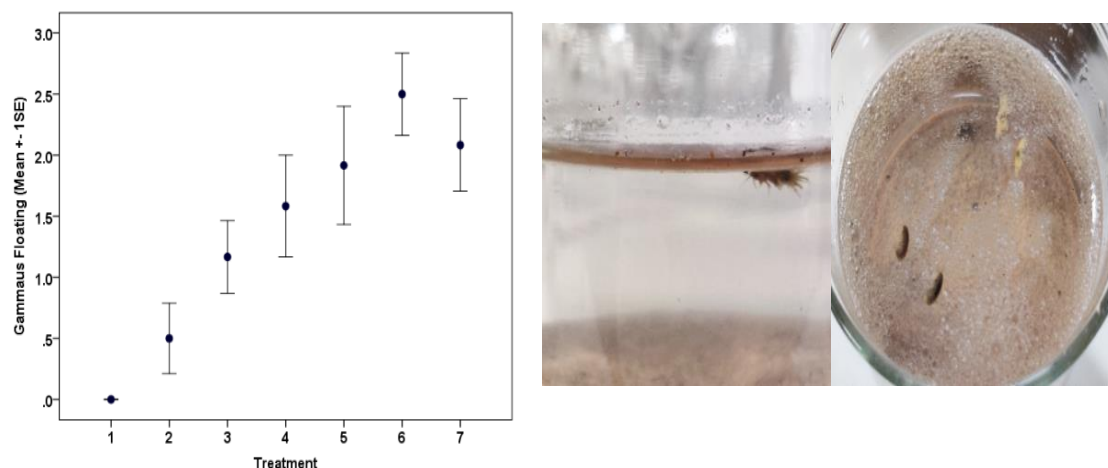


Figure 4: Mean number of *Gammarus pulex* (+/- 1 standard error) displaying floating behaviour in different treatments of mineral oil. Treatments 1-7 are mesocosm with increasing volumetric ratio of oil and water, with no mineral oil in Treatment 1 (control treatment), and 25 % in Treatment 7. As the quantity of oil increased, more invertebrates were active in the top surface layer and were more buoyant. On average, 2.5 out of 5 *Gammarids* displayed this behaviour in Treatment 6 (20% Oil). *Gammarids* can be seen close to the interface of the oil-water emulsion in photos on the right

This type of species response is significant in eco-toxicological studies where changes in behaviour are taken as endpoints of studies undertaking contaminant risk assessments (Peeters *et al.*, 2009). Although mortality is more typically the endpoint considered, the exposure to contamination at below-lethal concentrations may disturb and disrupt individuals' overall performance, affecting their long-term survival. The response may reflect the direct effect of environmental pollution (toxicosis) or the species' response mechanism(s) when exposed to the pollutant(s) (Peeters *et al.*, 2009). The response(s) recorded at the individual organism level are also important as they may help explain changes in the wider population and therefore, it is critical to establish associations between potential toxicity effects and ecosystem effects due to other factor such as behavioural change (Boyd *et al.*, 2002). Mesocosm studies, as illustrated above, help provide evidence required for developing larger scale monitoring programmes. Such laboratory scale investigations are critical to identify the appropriate biomonitoring tools and suitable organisms within specific geographical regions. The responses may aid in the development of both laboratory and field techniques with a view for the development of standard monitoring systems in the future (Delmail, 2014).

6. Future prospects of Integrated Monitoring in Low to Middle Income Countries (LMICs).

Reviewing contemporary soil-water pollution literature, with direct consideration of LMICs, indicates that conventional physico-chemical monitoring does not align with the rapidly evolving patterns of contamination from industrial, domestic, and agricultural sources. The ever-increasing range of emerging contaminants, potentially posing serious threats to ecosystems and environmental health, require urgent academic and regulatory authority attention. Biomonitoring provides a mechanism and range of techniques to characterise and quantify the effects of these substances which would otherwise be excluded from the list of potential threats due to a lack of evidence and data (Brack *et al.*, 2017). Ecosystem monitoring provides useful coverage of ecological effects and risks associated with chemical contamination. However, it does not discriminate between specific chemical stressors and other abiotic factors (e.g., climatic variation) which may also cause ecological variability. Therefore, it is essential to investigate the biological effects at an early-stage in order establish an association with chemical exposure and direct 'effects' (Milinkovitch *et al.*, 2019). Compartment-wise investigation of pollution

(water → sub-individual → individual → population) will provide the data required to identify the culpable contaminants and their environment fate and transport within the respective ecosystems.

The three principal types of monitoring methods applied in pollution investigations are illustrated in Figure 5. Physico-chemical and biological monitoring approaches have largely been applied independently from each other historically (Altenburger *et al.*, 2019). The authors argue that these methods provide maximum information when employed together. An integrated site monitoring framework as advocated in this paper would help address the current challenges and knowledge gaps. Enhanced efforts to incorporate bioaccumulation and toxicity monitoring within existing monitoring programmes will help identify the effects of non-targeted chemicals / emerging contaminants before effects are identified at higher levels (ecosystem) (Depledge *et al.*, 1995; Milinkovitch *et al.*, 2019) or they pose a threat to human health. The data generated via this holistic approach will provide greater information about exiting environmental resource quality and ecosystem services degradation.

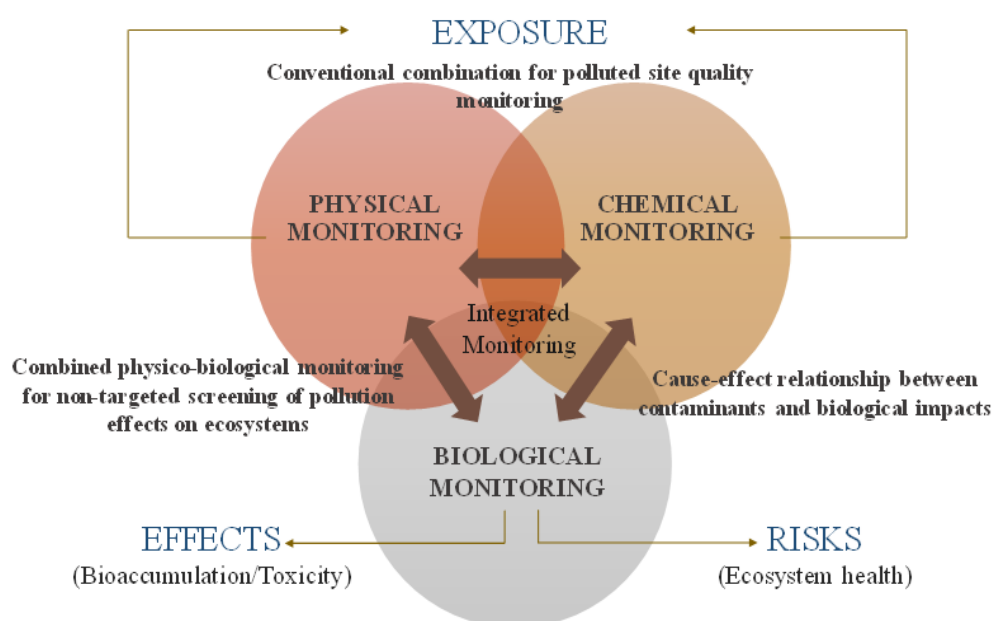


Figure 5: Interrelationships among physical, chemical and biological monitoring

A framework for integrated monitoring of polluted sites is outlined in Figure 6. The framework can be applied to investigate anthropogenically polluted sites using physical, chemical and biological parameters. It proposes a stage / longitudinal monitoring approach which progresses from the source location to receiving water bodies. Biomonitoring allows the collection of time-integrated information over extended period regarding pollutants which can account for seasonal and inter-annual variability in many instances. Utilising a framework such as proposed would establish planned monitoring programmes for the assessment of sites for chemical exposure, biological effects and wider risks to the ecosystem(s) and human health and may represent significant economic cost savings for many LMICs if implemented in a co-ordinated manner. Conventionally, the chemical and ecological status of polluted sites has required two independent assessments, typically by different agencies in low- and middle-income countries. This leads to duplication of effort and data as tests need to be completed for both types of assessments, resulting in increased costs to characterize the pollution at the same site. The integrated approach proposed would facilitate the identification of specific pollutants through chemical monitoring and also quantifying ecological damage and effects using biomonitoring techniques. However, instead of biologically monitoring each site subsequently, identification of key locations allows streamlining and prioritization of effort and resources by identifying the hot spots through effect-

based monitoring. This potentially reduces the overall cost by removing the need to monitor the entire network. The integrated approach is especially beneficial for sites subject to pollution mixtures with high chemical concentrations but limited human health impacts or ecosystem degradation. In some instances, the ecosystem or human health risks may be less than indicated by chemical characterization alone. It should be noted that in the short-term, the combined approach may require more sampling, although, in the long-term, financial savings may be made when an appropriate integrated monitoring system is practiced. This new system may also help to prevent future pollution by identifying potential problems/locations that can be remediated earlier at less cost than the long-term remediation if not addressed early. Thus, the costs of acquiring additional information by 1) avoiding duplication of effort/sampling, 2) streamlining pollution monitoring effects, and 3) developing networks that serve as early warning monitoring sites may prevent large-scale damage and identify problems before they escalate. Incorporating biological parameters allows the comparison of effects in various compartments of the receiving ecosystem / sites and at different biological levels. This also facilitates pollutant tracing from source to sink and bioaccumulation within food-chains and organisms at different trophic levels respectively. Some existing monitoring programs do not consider water bodies where pollutant concentrations are typically below detectable limits and focus on the acutely polluted/degraded sites. The authors suggest that these locations may be ideal for ecosystem monitoring as they may represent ideal reference (control sites), especially if they are in the vicinity of known polluted sites.

Sampling is a crucial step in accurately characterizing polluted sites. The optimal selection of sampling locations/sites, frequency, timing, sample size, volume, and pattern needs to reflect the investigation objectives and the properties of polluting substance and proposed sampling sites to be characterized (Namieśnik and Szefer, 2010). Water samples for physico-chemical analyses need to be taken from both upstream and downstream locations of effluent discharge points to provide a quantitative measure of the effect and a comparison with an unpolluted (control) site. Unrepresentative samples from the water body surface, bottom, boundaries/banks, and confluence zones of streams should be avoided (Namieśnik and Szefer, 2010). Samples from these locations present uncertainties due to their heterogeneity and the mixing properties of the water body. Biota sampling should be performed either through sampling specific locations or passive sampling devices (PSDs) depending on the contaminant type, habitat heterogeneity, and resource availability/sampling costs. Passive sampling should be undertaken for specific contaminants, including hydrophobic organic pollutants and polar substances, which display high concentration variability in aquatic environments (Brack *et al.*, 2017; Altenburger *et al.*, 2019). Integrating passive sampling methods in a time-integrated manner is suggested for quantification of hydrophobic/non-polar organic substances that frequently occur at trace level concentrations (El-Shenawy *et al.*, 2009; Miège *et al.*, 2015; Booij *et al.*, 2016). In addition, adsorption passive sampling should be undertaken to improve the representativeness of temporally variable hydrophilic/polar substances (Miège *et al.*, 2015; Brack *et al.*, 2017).

The proposed framework potentially provides multiple benefits if adopted more widely within LMICs, but also presents new challenges in the form of quantifying new and emerging pollutants and in the interpretation and analysis of the data. First, the effect(s) of confounding factors, including both intrinsic biotic (e.g., organisms size/weight/age, sex, and reproductive status) and extrinsic abiotic (e.g., pH, temperature, redox status, and salinity) factors needs to be considered to differentiate their effects from the pollutant stress under investigation. Second, the floral or faunal community level responses need to be undertaken on a long-term basis given that the effect of pollution may persist for multiple years and monitoring the post-event recovery may also be required. Third, the selection of biomarkers/bioindicators may be challenging in many LMIC regions where the taxonomy or many floral and faunal groups is not fully resolved, making implementation by inexperienced field practitioners even more challenging.

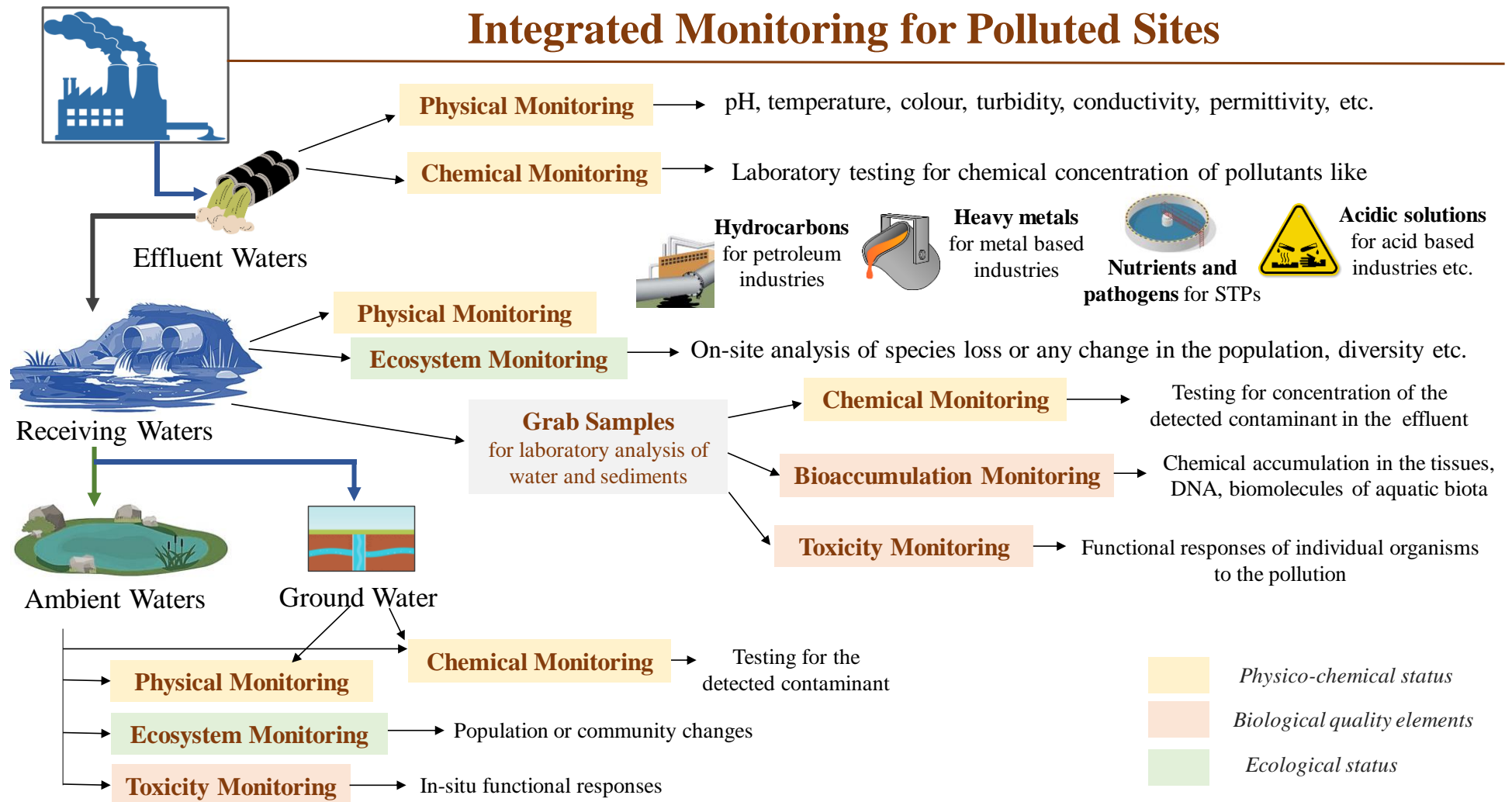


Figure 6: Schematic diagram presenting the framework for integrated monitoring of anthropogenically polluted sites

7. Conclusion

This review presents a framework for improving understanding and implementation of pollution monitoring and sampling strategies to reflect contemporary regulatory and research needs within Low to Middle Income Countries. The paper has identified knowledge gaps in the monitoring toolbox currently employed and has presented recommendations for comprehensive assessments within LMICs. Integration of biomonitoring approaches will help address and reduce the risks associated with transformation products, pollutant cocktails, and analytically undetected eco-toxic substances. This paper emphasizes the need to utilise information on chemical contamination and propagating its effects at various biological levels within the ecosystem and on human health. The effective integration of ‘effect – based’ and ‘risk – based’ monitoring will help improve monitoring of emerging and new (potentially unidentified) chemical and non-chemical stressors.

The review highlights the importance of the implementation and significance of the three biomonitoring methods that together form the basis for an integrated framework for monitoring anthropogenically polluted sites. Integrated monitoring, which is currently used to safeguard water resources (including hydrological, ecological, and societal resources) in many high-income countries, can be effectively applied for the assessment of polluted sites globally. While the current plans only consider ecosystem monitoring, the proposed integrated site monitoring framework should also incorporate biomarkers and bioassays to be most effective. This framework will help guide practitioners to adopt the most appropriate and cost – effective monitoring plans after selecting the ideal ‘sentinel’ (indicator) species and measurable parameters based on the local conditions. For real field technological application, economic and policy factors also play a critical role, and thus, the proposed integrated site monitoring framework will provide great benefits to LMICs by being cost effective and providing a readily transferable approach that allows the results from a polluted site to be considered in a wider ecosystem context.

The integrated approach outlined in this paper and the resulting conceptual framework make an important contribution to the environmental literature in terms of understanding the role of biological monitoring in pollution assessment in LMICs. Although the material presented focusses on soil-water pollution, the approach and framework could be readily transferred to other environments. Future research and assessments are required, especially within polluted soils, urban locations, and industrial/post-industrial ‘brownfield sites’ where the validation of the methods and framework would deliver maximum benefits.

Acknowledgements

The authors are thankful to Department of Science Technology, India and British Council, UK for joint support of the work through a UKIERI Project titled “In-situ bioremediation of non-aqueous phase liquids (NAPLs) pollution within the Baddi-Barotiwala Nalagarh (BBN) Industrial Area in Himachal Pradesh (India).” The first author is also thankful to the University Grants Commission (UGC) of India for providing support through a research fellowship. The last three authors acknowledge a travel grant provided by the Institute of Advanced Studies (IAS), Loughborough University, UK which enabled the 2nd author to make a trip to Loughborough and discuss the scope of this paper. All authors are thankful to NIH Roorkee, India, PSI Dehradun, India for their support to the above UKIERI project.

Declaration of competing interest: Authors declare no conflict of interest

References

- Adu-manu, K. S., Tapparello, C. and Heinzelman, W. (2017) ‘Soil Quality Monitoring using Wireless Sensor Network’, *International Journal of Recent Trends in Engineering and Research*, 3(8), pp. 218–222. doi: 10.23883/ijrter.2017.3407.bmrj0.
- Ali, S. *et al.* (2020) ‘Application of floating aquatic plants in phytoremediation of heavy metals polluted water:

47 A review', *Sustainability (Switzerland)*, 12(5), pp. 1–33. doi: 10.3390/su12051927.

48 Altenburger, R. *et al.* (2019) 'Future water quality monitoring: improving the balance between exposure and
49 toxicity assessments of real-world pollutant mixtures', *Environmental Sciences Europe*. S`pringer Berlin
50 Heidelberg, 31(1), pp. 1–17. doi: 10.1186/s12302-019-0193-1.

51 Altinok, I. and Capkin, E. (2007) 'Histopathology of Rainbow Trout Exposed to Sublethal Concentrations of
52 Methiocarb or Endosulfan', *Toxicologic Pathology*, 35(3), pp. 405–410. doi: 10.1080/01926230701230353.

53 Amiard-Triquet, C., Amiard, J.-C. and Mouneyrac, C. (2015) *Aquatic ecotoxicology: Advancing Tools for
54 Dealing with Emerging Risks*, Mica Haley. doi: 10.1007/BF02538101.

55 Amiard-Triquet, C., Amiard, J.-C. and Rainbow, P. S. (2013) *Ecological Biomarkers: Indicators of
56 Ecotoxicological Effects*, CRC Press, Taylor & Francis Group, Boca Raton.

57 Amiard-Triquet, C. and Rainbow, P. S. (2009) *Environmental Assessment of Ecosystems Estuarine: A Case
58 Study*. Taylor & Francis. doi: 10.1017/CBO9781107415324.004.

59 Artico, L. L., Migita, N. A. and Menezes, A. P. S. (2018) 'Toxicological Effects of Surface Water Exposed to
60 Coal Contamination on the Test System Allium cepa', *Water, Air, and Soil Pollution*. Water, Air, & Soil
61 Pollution, 229(248), pp. 1–12. doi: 10.1007/s11270-018-3904-0.

62 Baldantoni, D. *et al.* (2018) 'Biomonitoring of nutrient and toxic element concentrations in the Sarno River
63 through aquatic plants', *Ecotoxicology and Environmental Safety*. Elsevier Inc., 148(November 2017), pp. 520–
64 527. doi: 10.1016/j.ecoenv.2017.10.063.

65 Barnett-Itzhaki, Z. *et al.* (2018) 'A review of human biomonitoring in selected Southeast Asian countries',
66 *Environment International*. Elsevier, 116(March), pp. 156–164. doi: 10.1016/j.envint.2018.03.046.

67 Bartram, J. and Ballance, R. (1996) *Water Quality Monitoring - A Practical Guide to the Design and
68 Implementation of Freshwater Quality Studies and Monitoring Programmes*, Taylor and Francis, London and
69 New York.

70 Behmel, S. *et al.* (2016) 'Water quality monitoring strategies — A review and future perspectives', *Science of
71 the Total Environment*. Elsevier B.V., 571, pp. 1312–1329. doi: 10.1016/j.scitotenv.2016.06.235.

72 Beniah Obinna, I. and Ebere, *Enyoh Christian (2019) 'A review: Water pollution by heavy metal and organic
73 pollutants: Brief review of sources, effects and progress on remediation with aquatic plants', *Analytical Methods
74 in Environmental Chemistry Journal*, 2(3), pp. 5–38. doi: 10.24200/amecj.v2.i03.66.

75 Besse, J. P. *et al.* (2013) 'Caged Gammarus fossarum (Crustacea) as a robust tool for the characterization of
76 bioavailable contamination levels in continental waters: Towards the determination of threshold values', *Water
77 Research*, 47(2), pp. 650–660. doi: 10.1016/j.watres.2012.10.024.

78 Bilal, M. *et al.* (2016) 'Mutagenicity and cytotoxicity assessment of biodegraded textile effluent by Ca-alginate
79 encapsulated manganese peroxidase', *Biochemical Engineering Journal*. Elsevier B.V., 109, pp. 153–161. doi:
80 10.1016/j.bej.2016.01.020.

81 Birk, S. *et al.* (2012a) 'Three hundred ways to assess Europe's surface waters: An almost complete overview of
82 biological methods to implement the Water Framework Directive', *Ecological Indicators*. Elsevier Ltd, 18, pp.
83 31–41. doi: 10.1016/j.ecolind.2011.10.009.

84 Birk, S. *et al.* (2012b) 'Three hundred ways to assess Europe's surface waters: An almost complete overview of
85 biological methods to implement the Water Framework Directive', *Ecological Indicators*. Elsevier Ltd, 18, pp.
86 31–41. doi: 10.1016/j.ecolind.2011.10.009.

87 Bo, T. *et al.* (2017) 'Biomonitoring with macroinvertebrate communities in Italy: What happened to our past
88 and what is the future?', *Journal of Limnology*, 76(S1), pp. 21–28. doi: 10.4081/jlimnol.2016.1584.

89 Bonanno, G., Borg, J. A. and Di Martino, V. (2017) 'Levels of heavy metals in wetland and marine vascular
90 plants and their biomonitoring potential: A comparative assessment', *Science of the Total Environment*. Elsevier
91 B.V., 576, pp. 796–806. doi: 10.1016/j.scitotenv.2016.10.171.

92 Bonnail, E. *et al.* (2017) 'A novel approach for acid mine drainage pollution biomonitoring using rare earth
93 elements bioaccumulated in the freshwater clam Corbicula fluminea', *Journal of Hazardous Materials*. Elsevier
94 B.V., 338, pp. 466–471. doi: 10.1016/j.jhazmat.2017.05.052.

- Bonnail, E., Macías, F. and Osta, V. (2019) 'Ecological improvement assessment of a passive remediation technology for acid mine drainage: Water quality biomonitoring using bivalves', *Chemosphere*. Elsevier Ltd, 219, pp. 695–703. doi: 10.1016/j.chemosphere.2018.12.037.
- Booij, K. *et al.* (2016) 'Passive Sampling in Regulatory Chemical Monitoring of Nonpolar Organic Compounds in the Aquatic Environment', *Environmental Science and Technology*, 50(1), pp. 3–17. doi: 10.1021/acs.est.5b04050.
- Boyd, W. A., Brewer, S. K. and Williams, P. L. (2002) 'Altered Behaviour of Invertebrates Living in Polluted Environments', in *Behavioural Ecotoxicology*, pp. 293–336.
- Brack, W. *et al.* (2017) 'Towards the review of the European Union Water Framework management of chemical contamination in European surface water resources', *Science of the Total Environment*. Elsevier B.V., 576, pp. 720–737. doi: 10.1016/j.scitotenv.2016.10.104.
- Brainerd, E. and Menon, N. (2015) 'Religion and Health in Early Childhood: Evidence from South Asia', *Population and Development Review*, 41(3), pp. 439–463. doi: 10.1111/j.1728-4457.2015.00067.x.
- Bünemann, E. K. *et al.* (2018) 'Soil quality – A critical review', *Soil Biology and Biochemistry*. Elsevier, 120(September 2017), pp. 105–125. doi: 10.1016/j.soilbio.2018.01.030.
- Buss, D. F. *et al.* (2015) 'Stream biomonitoring using macroinvertebrates around the globe: a comparison of large-scale programs', *Environmental Monitoring and Assessment*, 187(1). doi: 10.1007/s10661-014-4132-8.
- Calvo-Flores, F. G., Isac-Garcia, J. and Dobado, J. A. (2018) *Emerging Pollutants, Handbook of Environmental Analysis*. doi: 10.1201/b10505-26.
- Carter, J. L. *et al.* (2006) 'Biomonitoring in North American Rivers: A Comparison of Methods Used for Benthic Macroinvertebrates in Canada and the United States', in Ziglio, G., Siligardi, M., and Flaim, G. (eds) *Water Quality Measurements Series*. John Wiley & Sons, Ltd, pp. 203–228.
- Carvalho, L. *et al.* (2019) 'Protecting and restoring Europe's waters: An analysis of the future development needs of the Water Framework Directive', *Science of the Total Environment*. The Authors, 658, pp. 1228–1238. doi: 10.1016/j.scitotenv.2018.12.255.
- Cavusoglu, Kultigin *et al.* (2010) 'Protective role of Ginkgo biloba on petroleum wastewater-induced toxicity in Vicia faba L. (Fabaceae) root tip cells', *Journal of Environmental Biology*, 31(3), pp. 319–324.
- Celander, M. C. (2011) 'Cocktail effects on biomarker responses in fish', *Aquatic Toxicology*. Elsevier B.V., 105(3-4 SUPPL.), pp. 72–77. doi: 10.1016/j.aquatox.2011.06.002.
- Chapman, P. M. (2002) 'Integrating toxicology and ecology: Putting the "eco" into ecotoxicology', *Marine Pollution Bulletin*, 44(1), pp. 7–15. doi: 10.1016/S0025-326X(01)00253-3.
- Ciliberti, A. *et al.* (2017) 'Caged Gammarus as biomonitors identifying thresholds of toxic metal bioavailability that affect gammarid densities at the French national scale', *Water Research*, 118, pp. 131–140. doi: 10.1016/j.watres.2017.04.031.
- Clark, J. L. and Clements, W. H. (2006) 'The use of in situ and stream microcosm experiments to assess population- and community-level responses to metals', *Environmental Toxicology and Chemistry*, 25(9), pp. 2306–2312. doi: 10.1897/05-552.1.
- Clarke, R. T. *et al.* (2006) 'Effects of sampling and sub-sampling variation using the STAR-AQEM sampling protocol on the precision of macroinvertebrate metrics', *Hydrobiologia*, 566(1), pp. 441–459. doi: 10.1007/s10750-006-0078-3.
- Costa, F. O. *et al.* (2005) 'Multi-level assessment of chronic toxicity of estuarine sediments with the amphipod Gammarus locusta: II. Organism and population-level endpoints', *Marine Environmental Research*, 60(1), pp. 93–110. doi: 10.1016/j.marenvres.2004.08.005.
- Dadea, C. *et al.* (2017) 'Tree species as tools for biomonitoring and phytoremediation in urban environments: A review with special regard to heavy metals', *Arboriculture and Urban Forestry*, 43(4), pp. 155–167. doi: 10.48044/auuf.2017.014.
- Dallas, H. F. (1997) 'A preliminary evaluation of aspects of sass (south african scoring system) for the rapid bioassessment of water quality in rivers, with particular reference to the incorporation of sass in a national

143 biomonitoring programme', *Southern African Journal of Aquatic Sciences*, 23(1), pp. 79–94. doi:
144 10.1080/10183469.1997.9631389.

145 Dallas, H. F. (2004) 'Seasonal variability of macroinvertebrate assemblages in two regions of South Africa:
146 Implications for aquatic bioassessment', *African Journal of Aquatic Science*, 29(2), pp. 173–184. doi:
147 10.2989/16085910409503808.

148 Dalu, T. and Froneman, P. W. (2016) 'Diatom-based water quality monitoring in southern Africa: Challenges
149 and future prospects', *Water SA*, 42(4), pp. 551–559. doi: 10.4314/wsa.v42i4.05.

150 Dauvin, J. C. and Ruellet, T. (2007) 'Polychaete/amphipod ratio revisited', *Marine Pollution Bulletin*, 55(1–6),
151 pp. 215–224. doi: 10.1016/j.marpolbul.2006.08.045.

152 Debén, S. *et al.* (2016) 'Using devitalized moss for active biomonitoring of water pollution', *Environmental*
153 *Pollution*, 210, pp. 315–322. doi: 10.1016/j.envpol.2016.01.009.

154 Debén, S. *et al.* (2017) 'Monitoring river water quality with transplanted bryophytes: A methodological review',
155 *Ecological Indicators*. Elsevier, 81(June), pp. 461–470. doi: 10.1016/j.ecolind.2017.06.014.

156 Delmail, D. (2014) 'Risk management of European inland waters using macrophyte biomonitoring', *Frontiers*
157 *in Environmental Science*, 2(JUL), pp. 2012–2015. doi: 10.3389/fenvs.2014.00031.

158 Depledge, M. H., Aagaard, A. and Györkös, P. (1995) 'Assessment of trace metal toxicity using molecular,
159 physiological and behavioural biomarkers', *Marine Pollution Bulletin*, 31(1–3), pp. 19–27. doi: 10.1016/0025-
160 326X(95)00006-9.

161 El-Shenawy, N. S. *et al.* (2009) 'Comparing the passive sampler and biomonitoring of organic pollutants in
162 water: A laboratory study', *Ocean Science Journal*, 44(2), pp. 69–77. doi: 10.1007/s12601-009-0008-1.

163 Farkas, A., Salánki, J. and Specziár, A. (2002) 'Relation between growth and the heavy metal concentration in
164 organs of bream *Abramis brama* L. populating Lake Balaton', *Archives of Environmental Contamination and*
165 *Toxicology*, 43(2), pp. 236–243. doi: 10.1007/s00244-002-1123-5.

166 Favas, P. J. C. *et al.* (2018) 'Metal(loid) accumulation in aquatic plants of a mining area: Potential for water
167 quality biomonitoring and biogeochemical prospecting', *Chemosphere*, 194, pp. 158–170. doi:
168 10.1016/j.chemosphere.2017.11.139.

169 Fu, Q. *et al.* (2018) 'Bioaccumulation, Biotransformation, and Synergistic Effects of Binary Fungicide Mixtures
170 in *Hyalella azteca* and *Gammarus pulex*: How Different/Similar are the Two Species?', *Environmental Science*
171 *and Technology*, 52(22), pp. 13491–13500. doi: 10.1021/acs.est.8b04057.

172 García-Seoane, R., Aboal, J. R., *et al.* (2018) 'Biomonitoring coastal environments with transplanted
173 macroalgae: A methodological review', *Marine Pollution Bulletin*. Elsevier, 135(February), pp. 988–999. doi:
174 10.1016/j.marpolbul.2018.08.027.

175 García-Seoane, R., Fernández, J. A., *et al.* (2018) 'Use of macroalgae to biomonitor pollutants in coastal waters:
176 Optimization of the methodology', *Ecological Indicators*. Elsevier, 84(September 2017), pp. 710–726. doi:
177 10.1016/j.ecolind.2017.09.015.

178 Gaskell, P. N., Brooks, A. C. and Maltby, L. (2007) 'Variation in the bioaccumulation of a sediment-sorbed
179 hydrophobic compound by benthic macroinvertebrates: Patterns and mechanisms', *Environmental Science and*
180 *Technology*, 41(5), pp. 1783–1789. doi: 10.1021/es061934b.

181 Gecheva, G. *et al.* (2015) 'Monitoring of aquatic mosses and sediments: a case study in contaminated rivers,
182 Bulgaria', *Plant Biosystems*. Taylor & Francis, pp. 527–536. doi: 10.1080/11263504.2013.857736.

183 Gerhardt, A. (1995) 'Monitoring behavioural responses to metals in *Gammarus pulex* (L.) (Crustacea) with
184 impedance conversion', *Environmental Science and Pollution Research*, 2(1), pp. 15–23. doi:
185 10.1007/BF02987506.

186 Gerhardt, A. *et al.* (2012) 'Active in Situ Biomonitoring of Pesticide Pulses Using *Gammarus* spp. in Small
187 Tributaries of Lake Constance', *Journal of Environmental Protection*, 03(07), pp. 573–583. doi:
188 10.4236/jep.2012.37069.

189 Gogoi, A. *et al.* (2018) 'Occurrence and fate of emerging contaminants in water environment: A review',
190 *Groundwater for Sustainable Development*. Elsevier, 6(September 2017), pp. 169–180. doi:

10.1016/j.gsd.2017.12.009.

Guareschi, S., Laini, A. and Sánchez-Montoya, M. M. (2017) 'How do low-abundance taxa affect river biomonitoring? Exploring the response of different macroinvertebrate-based indices', *Journal of Limnology*, 76(S1), pp. 9–20. doi: 10.4081/jlimnol.2016.1516.

Hale, S. L. and Koprowski, J. L. (2018) 'Ecosystem-level effects of keystone species reintroduction: a literature review', *Restoration Ecology*, 26(3), pp. 439–445. doi: 10.1111/rec.12684.

Hamza-Chaffai, A. (2014) 'Usefulness of Bioindicators and Biomarkers in Pollution Biomonitoring', *International Journal of Biotechnology for Wellness Industries*, 3(1), pp. 19–26. doi: 10.6000/1927-3037.2014.03.01.4.

Harmancioglu, N. B. *et al.* (1999) *Water Quality Monitoring Network Design*, Kluwer Academic Publishers, Dordrecht, the Netherlands. doi: 10.1007/0-306-48065-4.

Hering, D. *et al.* (2006) 'Cook book for the development of a Multimetric Index for biological condition of aquatic ecosystems: Experiences from the European AQEM and STAR projects and related initiatives', *Hydrobiologia*, 566(1), pp. 311–324. doi: 10.1007/s10750-006-0087-2.

Huang, Y. *et al.* (2019) 'Current status of agricultural soil pollution by heavy metals in China: A meta-analysis', *Science of the Total Environment*. Elsevier B.V., 651, pp. 3034–3042. doi: 10.1016/j.scitotenv.2018.10.185.

Hybská, H. *et al.* (2020) 'Biomonitoring and its in the assessment of the quality of wastewater treatment process', *Environmental Nanotechnology, Monitoring and Management*. Elsevier, 13(March), p. 100292. doi: 10.1016/j.enmm.2020.100292.

Ighalo, J. O. and Adeniyi, A. G. (2020) 'A comprehensive review of water quality monitoring and assessment in Nigeria', *Chemosphere*. Elsevier Ltd, 260, p. 127569. doi: 10.1016/j.chemosphere.2020.127569.

Janssens De Bisthoven, L., Vermeulen, A. and Ollevier, F. (1998) 'Experimental induction of morphological deformities in *Chironomus riparius* larvae by chronic exposure to copper and lead', *Archives of Environmental Contamination and Toxicology*, 35(2), pp. 249–256. doi: 10.1007/s002449900373.

Japitana, M. V. *et al.* (2018) 'Integrated technologies for low cost environmental monitoring in the water bodies of the Philippines: A review', *Nature Environment and Pollution Technology*, 17(4), pp. 1125–1137.

Jemec, A. *et al.* (2008) 'Biochemical biomarkers in chronically metal-stressed daphnids', *Comparative Biochemistry and Physiology - C Toxicology and Pharmacology*, 147(1), pp. 61–68. doi: 10.1016/j.cbpc.2007.07.006.

Jemec, A. *et al.* (2010) 'Biochemical biomarkers in environmental studies-lessons learnt from enzymes catalase, glutathione S-transferase and cholinesterase in two crustacean species', *Environmental Science and Pollution Research*, 17(3), pp. 571–581. doi: 10.1007/s11356-009-0112-x.

Kapfer, J. *et al.* (2012) 'Do bryophytes show a stronger response than vascular plants to interannual changes in spring water quality?', *Freshwater Science*, 31(2), pp. 625–635. doi: 10.1899/11-037.1.

Kunz, P. Y., Kienle, C. and Gerhardt, A. (2010) *Gammarus spp. in aquatic ecotoxicology and water quality assessment: Toward integrated multilevel tests*, *Reviews of Environmental Contamination and Toxicology*. doi: 10.1007/978-1-4419-5623-1_1.

Lavaine, E. (2015) 'An Econometric Analysis of Atmospheric Pollution, Environmental Disparities and Mortality Rates', *Environmental and Resource Economics*, 60(2), pp. 215–242. doi: 10.1007/s10640-014-9765-0.

Lebrun, J. D. *et al.* (2015) 'Seasonal variability and inter-species comparison of metal bioaccumulation in caged gammarids under urban diffuse contamination gradient: Implications for biomonitoring investigations', *Science of the Total Environment*. Elsevier B.V., 511, pp. 501–508. doi: 10.1016/j.scitotenv.2014.12.078.

Lebrun, J. D. and Gismondi, E. (2020) 'Behavioural and biochemical alterations in gammarids as induced by chronic metallic exposures (Cd, Cu and Pb): Implications for freshwater biomonitoring', *Chemosphere*. Elsevier Ltd, 257, p. 127253. doi: 10.1016/j.chemosphere.2020.127253.

Lee, W. Y. and Wang, W. X. (2001) 'Metal accumulation in the green macroalga *Ulva fasciata*: Effects of nitrate, ammonium and phosphate', *Science of the Total Environment*, 278(1–3), pp. 11–22. doi:

239 10.1016/S0048-9697(00)00884-6.

240 Li, C. *et al.* (2018) 'An integrated approach with the zebrafish model for biomonitoring of municipal wastewater
241 effluent and receiving waters', *Water Research*. Elsevier Ltd, 131, pp. 33–44. doi:
242 10.1016/j.watres.2017.12.017.

243 Li, L., Zheng, B. and Liu, L. (2010) 'Biomonitoring and bioindicators used for river ecosystems: Definitions,
244 approaches and trends', *Procedia Environmental Sciences*, 2(July), pp. 1510–1524. doi:
245 10.1016/j.proenv.2010.10.164.

246 Liu, W. X. *et al.* (2016) 'Aquatic biota as potential biological indicators of the contamination, bioaccumulation
247 and health risks caused by organochlorine pesticides in a large, shallow Chinese lake (Lake Chaohu)',
248 *Ecological Indicators*. Elsevier Ltd, 60(May 2009), pp. 335–345. doi: 10.1016/j.ecolind.2015.06.026.

249 Lovett-doust, L. and Lovett-doust, J. (2001) 'Plant Biomonitoring in Aquatic Environments', *Biomonitoring and*
250 *Biomarkers as Indicators of Environmental Change* 2, (January), pp. 347–360. doi: 10.1007/978-1-4615-1305-
251 6.

252 Lovett Doust, J., Schmidt, M. and Lovett Doust, L. (1994) 'Biological assessment of aquatic pollution: A
253 review, with emphasis on plants as biomonitoring', *Biological Reviews of the Cambridge Philosophical Society*,
254 69(2), pp. 147–186. doi: 10.1111/j.1469-185X.1994.tb01504.x.

255 Mangadze, T., Dalu, T. and William Froneman, P. (2019) 'Biological monitoring in southern Africa: A review
256 of the current status, challenges and future prospects', *Science of the Total Environment*, 648(January), pp.
257 1492–1499. doi: 10.1016/j.scitotenv.2018.08.252.

258 Martin, S. and Griswold, W. (2009) 'Human health effects of heavy metals', *Center for Hazardous Substance*
259 ..., (15), pp. 1–6. Available at:
260 http://drupal.engg.ksu.edu/chsr/outreach/resources/docs/15_HumanHealthEffectsofHeavyMetals2013.pdf%5Cn
261 <http://www.engg.ksu.edu/chsr/files/chsr/outreach-resources/15HumanHealthEffectsofHeavyMetals.pdf>.

262 Mathers, K. L. *et al.* (2016) 'The implications of an invasive species on the reliability of macroinvertebrate
263 biomonitoring tools used in freshwater ecological assessments', *Ecological Indicators*, 63, pp. 23–28.

264 Mathur, S. and Yadav, B. K. (2009) 'Phytoextraction modeling of heavy metal (Lead) contaminated site using
265 maize (*Zea Mays*)', *Practice Periodical of Hazardous, Toxic, and Radioactive Waste Management*, 13(4), pp.
266 229–238. doi: 10.1061/(ASCE)1090-025X(2009)13:4(229).

267 McBratney, A., Field, D. J. and Koch, A. (2014) 'The dimensions of soil security', *Geoderma*. Elsevier B.V.,
268 213, pp. 203–213. doi: 10.1016/j.geoderma.2013.08.013.

269 Meador, J. P. *et al.* (1995) 'Bioaccumulation of polycyclic aromatic hydrocarbons by marine organisms.',
270 *Reviews of environmental contamination and toxicology*, 143, pp. 79–165. doi: 10.1007/978-1-4612-2542-3_4.

271 Melville, F. and Pulkownik, A. (2007) 'Investigation of mangrove macroalgae as biomonitoring of estuarine metal
272 contamination', *Science of the Total Environment*, 387(1–3), pp. 301–309. doi: 10.1016/j.scitotenv.2007.06.036.

273 Metcalfe-Smith, J. L. (1994) 'Biological Water-Quality Assessment of Rivers: Use of Macroinvertebrate
274 Communities', in *The Rivers Handbook: Hydrological and Ecological Principles*, pp. 144–170. doi:
275 10.1002/9781444313871.ch8.

276 Miège, C. *et al.* (2015) 'Position paper on passive sampling techniques for the monitoring of contaminants in the
277 aquatic environment - Achievements to date and perspectives', *Trends in Environmental Analytical Chemistry*,
278 8, pp. 20–26. doi: 10.1016/j.teac.2015.07.001.

279 Milinkovitch, T. *et al.* (2019) 'Biomarkers as tools for monitoring within the Water Framework Directive
280 context: concept, opinions and advancement of expertise', *Environmental Science and Pollution Research*.
281 Environmental Science and Pollution Research, 26(32), pp. 32759–32763. doi: 10.1007/s11356-019-06434-x.

282 Moe, S. J. *et al.* (2013) 'Combined and interactive effects of global climate change and toxicants on populations
283 and communities', *Environmental Toxicology and Chemistry*, 32(1), pp. 49–61. doi: 10.1002/etc.2045.

284 Mohammadzadeh Pakdel, P. and Peighambari, S. J. (2018) 'A review on acrylic based hydrogels and their
285 applications in wastewater treatment', *Journal of Environmental Management*. Elsevier Ltd, 217, pp. 123–143.
286 doi: 10.1016/j.jenvman.2018.03.076.

- 287 Morse, J. C. *et al.* (2007) 'Freshwater biomonitoring with macroinvertebrates in East Asia', *Frontiers in*
288 *Ecology and the Environment*, 5(1), pp. 33–42. doi: 10.1890/1540-9295(2007)5[33:FBWMIE]2.0.CO;2.
- 289 Namieśnik, J. and Szefer, P. (2010) *Analytical Measurements in Aquatic Environments*. CRC Press, Taylor &
290 Francis Group.
- 291 Nascimento, F. X., Rossi, M. J. and Glick, B. R. (2018) 'Ethylene and 1-aminocyclopropane-1-carboxylate
292 (ACC) in plant–bacterial interactions', *Frontiers in Plant Science*, 9(February), pp. 1–17. doi:
293 10.3389/fpls.2018.00114.
- 294 Neuparth, T. *et al.* (2005) 'Multi-level assessment of chronic toxicity of estuarine sediments with the amphipod
295 *Gammarus locusta*: I. Biochemical endpoints', *Marine Environmental Research*, 60(1), pp. 69–91. doi:
296 10.1016/j.marenvres.2004.08.006.
- 297 Niba, A. and Sakwe, S. (2018) 'Turnover of benthic macroinvertebrates along the Mthatha River, Eastern Cape,
298 South Africa: implications for water quality bio-monitoring using indicator species', *Journal of Freshwater*
299 *Ecology*. Taylor & Francis, 33(1), pp. 157–171. doi: 10.1080/02705060.2018.1431969.
- 300 Odountan, O. H. *et al.* (2019) 'Biomonitoring of lakes using macroinvertebrates: recommended indices and
301 metrics for use in West Africa and developing countries', *Hydrobiologia*. Springer International Publishing,
302 826(1). doi: 10.1007/s10750-018-3745-2.
- 303 Von Der Ohe, P. C. and Liess, M. (2004) 'Relative sensitivity distribution of aquatic invertebrates to organic
304 and metal compounds', *Environmental Toxicology and Chemistry*, 23(1), pp. 150–156. doi: 10.1897/02-577.
- 305 Ollis, D. J. *et al.* (2006) 'Bioassessment of the ecological integrity of river ecosystems using aquatic
306 macroinvertebrates: An overview with a focus on South Africa', *African Journal of Aquatic Science*, 31(2), pp.
307 205–227. doi: 10.2989/16085910609503892.
- 308 Olusola, O. I. and Solomon, N. U. (2018) 'Genotoxicity assessment of three industrial effluents using the
309 *Allium cepa* bioassay', *African Journal of Environmental Science and Technology*, 12(3), pp. 115–122. doi:
310 10.5897/ajest2017.2447.
- 311 Oost, R. van der, Beyer, J. and Vermeulen, N. P. E. (2016) 'Soil Quality Field Kit : Part II', *Environmental*
312 *Toxicology and Pharmacology*, 13(February), pp. 57–149.
- 313 Paulino, M. G. *et al.* (2014) 'The impact of organochlorines and metals on wild fish living in a tropical
314 hydroelectric reservoir: Bioaccumulation and histopathological biomarkers', *Science of the Total Environment*.
315 Elsevier B.V., 497–498, pp. 293–306. doi: 10.1016/j.scitotenv.2014.07.122.
- 316 Peeters, E. T. H. M., De Lange, H. J. and Lurling, M. (2009) 'Variation in the behavior of the amphipod
317 *gammarus pulex*', *Human and Ecological Risk Assessment*, 15(1), pp. 41–52. doi:
318 10.1080/10807030802615055.
- 319 Phillips, D. J. H. and Rainbow, P. S. (1994) *Biomonitoring of trace aquatic contaminants*, Chapman & Hall.
320 Available at:
321 [http://apps.webofknowledge.com/full_record.do?product=UA&search_mode=GeneralSearch&qid=123&SID=V](http://apps.webofknowledge.com/full_record.do?product=UA&search_mode=GeneralSearch&qid=123&SID=V2M3ByOuOWpiZZWCxvo&page=1&doc=2)
322 [2M3ByOuOWpiZZWCxvo&page=1&doc=2](http://apps.webofknowledge.com/full_record.do?product=UA&search_mode=GeneralSearch&qid=123&SID=V2M3ByOuOWpiZZWCxvo&page=1&doc=2).
- 323 Piña, B. and Barata, C. (2011) 'A genomic and ecotoxicological perspective of DNA array studies in aquatic
324 environmental risk assessment', *Aquatic Toxicology*, 105(3-4 SUPPL.), pp. 40–49. doi:
325 10.1016/j.aquatox.2011.06.006.
- 326 Prabhakaran, K. *et al.* (2017) 'Biomonitoring of Malaysian aquatic environments: A review of status and
327 prospects', *Ecohydrology and Hydrobiology*. European Regional Centre for Ecohydrology of the Polish
328 Academy of Sciences, 17(2), pp. 134–147. doi: 10.1016/j.ecohyd.2017.03.001.
- 329 Pratas, J. *et al.* (2017) 'Distribution of rare earth elements, thorium and uranium in streams and aquatic mosses
330 of Central Portugal', *Environmental Earth Sciences*, 76(4). doi: 10.1007/s12665-017-6459-2.
- 331 Rainbow, P. S. (1995) 'Biomonitoring of heavy metal availability in the marine environment', *Marine Pollution*
332 *Bulletin*, 31(4–12), pp. 183–192. doi: 10.1016/0025-326X(95)00116-5.
- 333 Romero, J. *et al.* (2006) 'Nutrients Dynamics in Seagrass Ecosystems', in Larkum, A. W. D., Orth, R. J., and
334 Duarte, C. M. (eds) *Seagrasses: Biology, Ecology and Conservation*. Springer Berlin Heidelberg, pp. 227–254.

335 Salánki, J. *et al.* (2003) 'Molluscs in biological monitoring of water quality', *Toxicology Letters*, 140–141, pp.
336 403–410. doi: 10.1016/S0378-4274(03)00036-5.

337 Samsi, N. *et al.* (2017) 'Gastropods As A Bioindicator and Biomonitoring Water Pollution', *Aquacultura*
338 *Indonesiana*, 18(1), p. 54. doi: 10.21534/ai.v18i1.42.

339 Saunier, J. B. *et al.* (2013) 'Trace elements biomonitoring in a historical mining district (les Malines, France)',
340 *Chemosphere*. Elsevier Ltd, 93(9), pp. 2016–2023. doi: 10.1016/j.chemosphere.2013.07.024.

341 El Sayed, S. M. *et al.* (2020) 'An integrated water quality assessment of Damietta and Rosetta branches (Nile
342 River, Egypt) using chemical and biological indices', *Environmental Monitoring and Assessment*, 192(4). doi:
343 10.1007/s10661-020-8195-4.

344 Schiedek, D. *et al.* (2007) 'Interactions between climate change and contaminants', *Marine Pollution Bulletin*,
345 54(12), pp. 1845–1856. doi: 10.1016/j.marpolbul.2007.09.020.

346 Schilderman, P. A. E. L. *et al.* (1999) 'Use of crayfish in biomonitoring studies of environmental pollution of
347 the river Meuse', *Ecotoxicology and Environmental Safety*, 44(3), pp. 241–252. doi: 10.1006/eesa.1999.1827.

348 Schöne, B. R. and Krause, R. A. (2016) 'Retrospective environmental biomonitoring – Mussel Watch
349 expanded', *Global and Planetary Change*. Elsevier B.V., 144, pp. 228–251. doi:
350 10.1016/j.gloplacha.2016.08.002.

351 Serdar, O. *et al.* (2019) 'Modelling cadmium bioaccumulation in *Gammarus pulex* by using experimental design
352 approach', *Chemistry and Ecology*. Taylor & Francis, 35(10), pp. 922–936. doi:
353 10.1080/02757540.2019.1670814.

354 Serdar, O. (2019) 'The effect of dimethoate pesticide on some biochemical biomarkers in *Gammarus pulex*',
355 *Environmental Science and Pollution Research*, 26, pp. 21906–21914. doi: 10.1007/s11682-018-9832-1.

356 Shahid, N. *et al.* (2018) 'Adaptation of *Gammarus pulex* to agricultural insecticide contamination in streams',
357 *Science of the Total Environment*. Elsevier B.V., 621, pp. 479–485. doi: 10.1016/j.scitotenv.2017.11.220.

358 Tatar, S. *et al.* (2018) 'The using of *Gammarus pulex* as a biomonitor in ecological risk assessment of secondary
359 effluent from municipal wastewater treatment plant in Tunceli, Turkey', *Human and Ecological Risk*
360 *Assessment*. Taylor & Francis, 24(3), pp. 819–829. doi: 10.1080/10807039.2017.1400374.

361 Thrush, S. F. *et al.* (2008) 'Multiple stressor effects identified from species abundance distributions: Interactions
362 between urban contaminants and species habitat relationships', *Journal of Experimental Marine Biology and*
363 *Ecology*. Elsevier B.V., 366(1–2), pp. 160–168. doi: 10.1016/j.jembe.2008.07.020.

364 USEPA (2005) *Monitoring Data: Exploring your data, the first step*. Available at:
365 <https://www.epa.gov/polluted-runoff->.

366 Vanderpoorten, A. (1999) 'Aquatic bryophytes for a spatio-temporal monitoring of the water pollution of the
367 rivers Meuse and Sambre (Belgium)', *Environmental Pollution*, 104(3), pp. 401–410. doi: 10.1016/S0269-
368 7491(98)00170-5.

369 Vethaak, A. D. *et al.* (2017) 'Integrated indicator framework and methodology for monitoring and assessment of
370 hazardous substances and their effects in the marine environment', *Marine Environmental Research*, 124, pp.
371 11–20. doi: 10.1016/j.marenvres.2015.09.010.

372 Vieira, C. E. D. *et al.* (2019) 'An integrated approach in subtropical agro-ecosystems: Active biomonitoring,
373 environmental contaminants, bioaccumulation, and multiple biomarkers in fish', *Science of the Total*
374 *Environment*, 666, pp. 508–524. doi: 10.1016/j.scitotenv.2019.02.209.

375 Villares, R., Puente, X. and Carballeira, A. (2001) 'Ulva and Enteromorpha as indicators of heavy metal
376 pollution', *Hydrobiologia*, 462, pp. 221–232. doi: 10.1023/A.

377 Vincent, C. D., Lawlor, A. J. and Tipping, E. (2001) 'Accumulation of Al, Mn, Fe, Cu, Zn, Cd and Pb by the
378 bryophyte *Scapania undulata* in three upland waters of different pH', *Environmental Pollution*, 114(1), pp. 93–
379 100. doi: 10.1016/S0269-7491(00)00201-3.

380 Wang, Q. and Yang, Z. (2016a) 'Industrial water pollution, water environment treatment, and health risks in
381 China', *Environmental Pollution*. Elsevier Ltd, 218(November), pp. 358–365. doi:
382 10.1016/j.envpol.2016.07.011.

383 Wang, Q. and Yang, Z. (2016b) 'Industrial water pollution, water environment treatment, and health risks in
384 China', *Environmental Pollution*. Elsevier Ltd, 218, pp. 358–365. doi: 10.1016/j.envpol.2016.07.011.

385 Winkelbauer, A. *et al.* (2014) 'Crucial elements and technical implementation of intelligent monitoring
386 networks', *Water Science and Technology*, 70(12), pp. 1926–1933. doi: 10.2166/wst.2014.415.

387 Winkler, S. *et al.* (2008) 'Intelligent monitoring networks - Transformation of data into information for water
388 management', *Water Science and Technology*, 58(2), pp. 317–322. doi: 10.2166/wst.2008.672.

389 World Health Organization (2003) 'Anesthetic issues', in *Guidelines for Safe Recreational Water Environments*,
390 pp. 159–167. doi: 10.1201/b14143-5.

391 Yadav, B. K., Siebel, M. A. and van Bruggen, J. J. A. (2011) 'Rhizofiltration of a Heavy Metal (Lead)
392 Containing Wastewater Using the Wetland Plant *Carex pendula*', *Clean - Soil, Air, Water*, 39(5), pp. 467–474.
393 doi: 10.1002/clen.201000385.

394 Zalewski, M. (2015) 'Process oriented thinking as a key for integration of ecohydrology, biotechnology and
395 engineering for sustainable water resources management and ecosystems', *IAHS-AISH Proceedings and
396 Reports*, 366(June 2014), p. 187. doi: 10.5194/piahs-366-187-2015.

397 Zhang, C. *et al.* (2020) 'Microplastic pollution in surface water from east coastal areas of Guangdong, South
398 China and preliminary study on microplastics biomonitoring using two marine fish', *Chemosphere*. Elsevier
399 Ltd, 256, p. 127202. doi: 10.1016/j.chemosphere.2020.127202.

400 Zhang, T. *et al.* (2020) 'Study on the removal of aesthetic indicators by ozone during advanced treatment of
401 water reuse', *Journal of Water Process Engineering*. Elsevier, 36(April), pp. 1–7. doi:
402 10.1016/j.jwpe.2020.101381.

403 Zhou, Q. *et al.* (2008) 'Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic
404 ecosystem', *Analytica Chimica Acta*, 606(2), pp. 135–150. doi: 10.1016/j.aca.2007.11.018.

405 Zuykov, M., Pelletier, E. and Harper, D. A. T. (2013) 'Bivalve mollusks in metal pollution studies: From
406 bioaccumulation to biomonitoring', *Chemosphere*. Elsevier Ltd, 93(2), pp. 201–208. doi:
407 10.1016/j.chemosphere.2013.05.001.

408 Zwart, D. De (1995) 'Monitoring Water Quality in the Future Volume 3 : Biomonitoring', *National Institute of
409 Public Health & Environmental Protection (RIVM)*, 3.

410